VOL 33 (4) 2022: 566-574 | RESEARCH ARTICLE

Analgesic and Anti-inflammatory Activities of *Urena lobata* L. Leaf Extracts

Yudi Purnomo*, Andri Tilaqza

Department of Pharmacy, Faculty of Medicine, University of Islam, Malang 65411, Indonesia

ABSTRACT

Submitted: 21-07-2021 **Revised:** 02-02-2022 **Accepted:** 20-07-2022

*Corresponding author Yudi Purnomo

Email:

y_purnomo92@yahoo.com

Pulutan (Urena lobata L.) is a medicinal plant that has been traditionally used to treat fever, infection, and inflammation. However, preclinical studies on its activity as an analgesic and anti-inflammatory agent are still limited. The present study was aimed at investigating the analgesic and anti-inflammatory activities of *U. lobata* leaf extracts, as well as predicting the biological properties of the active components. Eight (two control and six test) groups of male Wistar rats were used in this study. Water and ethanol were used to extract the leaves of *U. lobata*, and each prepared in doses of 125, 250, and 500 mg/kg body weight (BW) for the treatments. The antiinflammatory and analgesic activities of the extracts were evaluated. Following active compound identification using UHPLC-MS/MS, several compounds were analyzed for drug-likeness and ADME-T properties. The results revealed that both aqueous and ethanolic extracts of *U. lobata* at 125 and 250 mg/kg BW significantly (p < 0.05) inhibited paw edema with an area under curve (AUC) volume of 10 and 5%, respectively, compared to the control group. For analgesic activity, the aqueous extract of *U. lobata* was able to significantly (p<0.05) increase the AUC for pain threshold by 30-100 % compared to the control group. However, the activity of the ethanolic extract was lower than the aqueous extract. The drug-likeness analysis indicated that all the phytoconstituents were within the range established by Lipinski's rule of five. Stigmasterol, \(\mathcal{B} \)-sitosterol, and mangiferin were predicted to have better ADME-T properties than that of the other compounds.

Keywords: Analgesic activity, Inflammation, Phytoconstituents, Pulutan, *Urena lobata*

INTRODUCTION

Inflammation is the underlying etiology of both degenerative and non-degenerative diseases. An inflammatory reaction is a physiological response of the body to protect itself against physical trauma, chemical agents, and microbial infection. During inflammation, some proinflammatory mediators such as prostaglandins, histamine, and leukotrienes are released, which causes the appearance of the inflammation cardinal signs: dolor (pain), tumor (swelling), rubor (red), calor (heat). Inflammatory cells like macrophages express the COX-2 enzyme, which catalyzes the conversion of arachidonic acid to prostaglandins (PGs) (Zambre et al., 2007). Inflammation plays a role in self-defense against injury factors. However, an uncontrolled and chronic inflammatory response would have a negative impact in form of hypersensitivity,

autoimmune and degenerative diseases (Price, 2006).

Opioid analgesics, steroids, and nonsteroidal anti-inflammatory drugs are used to treat both pain and inflammation, despite their adverse drug reactions. Medicinal plants can be used as an alternative source to substitute for them. Some herbs have been screened for their potential analgesic and anti-inflammatory activity. However, only a few of them have been reported. Therefore, exploring herbs to determine their active compounds would contribute to the current scientific knowledge (Sen *et al.*, 2010).

Pulutan (*Urena lobate* L.), belonging to the Malvaceae family, is a plant that has medicinal properties. It has been discovered that *U. lobata* ethanolic extract has anti-inflammatory properties (Babu *et al.*, 2016). According to preclinical research, active ingredients of *U. lobata*, such as

conditions of Creative Commons Attribution 2.0 Generic License (https://creativecommons.org/licenses/by/2.0/).

quercetin and mangiferin, have antioxidant, antibacterial, and diabetes-related properties (Adewale *et al.*, 2007; Purnomo *et al.*, 2021). Flavonoids and terpenoids are the main active compounds in herbs that have activity as an anti-inflammatory. Meanwhile, some alkaloids have analgesic and anti-inflammatory activities (Rivera *et al.*, 2006). Investigations into the analgesic and anti-inflammatory activities of *U. lobata* have not been fully explored. To promote the development and discovery of oral medications, it is necessary to study these bioactivities as well as the pharmacokinetic profile and toxicity predictions of the active compounds in *U. lobata*.

Therefore, the present study was conducted to examine the bioactivity, pharmacokinetic profile, and toxicity predictions of the active substances in *U. lobate* to support the development and discovery of drugs taken orally.

MATERIALS AND METHODS

Source of plant material

The *Urena lobata* leaf powder was obtained from Balai Materia Medika Batu Malang, Indonesia. The powder was identified and authenticated in the UPT Laboratorium Herbal Materia Medika Batu Malang with an ID number 074/027/101.8/2015.

Source of animal

The Wistar rats (*Rattus norvegicus*) used for the study were obtained from the Pusat Veteriner Farma (Pusvetma) Surabaya.

Ethical approval

Ethical approval for this study was obtained from the Commission of Ethical Research at Brawijaya University, Malang, Indonesia with approval ID number 002-KEP-UB-2020.

Preparation of plant extract

Extraction from *Urena lobata* leaf powder was carried out by boiling 50 g of leaf powder in 250 ml of hot water at 90°C for 30 minutes. The same quantity of *U. lobata* leaf powder was digested in 250 ml of ethanol for six hours using a water bath shaker, and the process was repeated three times with different solvents. A rotary evaporator was used to condense both extracts at the end.

Identification of active substances in Urena lobata leaf extracts

UHPLC (ACCELLA 1250, Thermo Scientific) and the detector, MS/MS Triple Q (Thermo Finnigan), were used in a qualitative analysis to

identify the active components from both the aqueous and ethanolic extracts of *U. lobata* leaves. The mobile phase used was 0.1 % formic acid in methanol and water. Ten active molecules from the phytosterol, flavonoid, and alkaloid families were identified.

Prediction of biological activities

The structures of the identified compounds were obtained from the PubChem database (https://pubchem.ncbi.nlm.nih.gov/). Using the Swiss ADME website (http://www.swissadme.ch/) for drug-likeness analysis, the physicochemical characteristics of Lipinski's rule of five were investigated. The pharmacokinetic properties (absorption, distribution, metabolism, excretion, and toxicity [ADME-T]) were analyzed using the pkCSM website (https://biosig.unimelb.edu.au/pkcsm).

Animal groupings and treatments

An *in vivo* experiment was conducted using male Wistar rats (180-200 g). The rats were divided into eight groups, each consisting of four rats. Within those eight groups, two control and six test groups were created. Each received an oral dose of 125, 250, and 500 mg/kg body weight (BW) of *U. lobata* aqueous extract (AUL) or ethanolic extract (EUL), respectively.

Anti-inflammatory activity assay

The extracts were administered orally an hour before testing. Following intraplanar injection of 1% carrageenan, anti-inflammatory activity was examined, and paw edema was monitored using a plethysmometer 6 h later. The data were presented as area under curve (AUC) of paw edema using ibuprofen 20 mg/kg as a reference drug.

Analgesic activity assay

One hour before the test, the extracts were administered orally. Analgesic activity was examined by an analgesymeter for 4h. Ibuprofen (20 mg/kg p.o.) was used as a reference drug while the pain threshold was being monitored. An AUC of pain threshold was used to display the data.

Statistical analysis

The data were presented as mean \pm SD. Statistical analysis was performed by one-way ANOVA (SPSS version 25). Statistical significance was determined using the least significant difference (LSD) test and p < 0.05 was considered statistically significant.

RESULTS AND DISCUSSION

Bioactive substances present in Urena lobata leaf extracts

According to the results (Figure 1 and Table II) of the UHPLC-MS/MS analysis of the aqueous and ethanolic extracts of $\it U.\ lobata$ leaves, the bioactive compounds were stigmasterol, gossypetin, and $\it \beta$ -sitosterol with a lesser concentration of mangiferin and chrysoeriol.

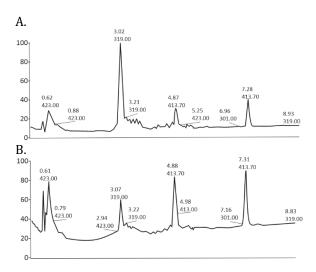


Figure 1. Chromatogram of active compound *U. lobata* (A). Water extract (B). Ethanolic extract

Predicted bioactivity of Urena lobata leaf extracts

The physicochemical characteristics and pharmacokinetics of the active ingredient from *U*. lobata as determined by an in silico analysis (Table II and III). To determine whether a molecule has the potential to be developed into a medication, Lipinski's rule of five is applied (drug-likeness). This rule predicts that a molecule has a high likelihood of being developed into a drug if there are no more than two exceptions to the following conditions: there are no more than five H-bond donors (HBD) and 10 H-bond acceptors (HBA), the molecular weight (MW) is smaller than 500, and the calculated LogP is smaller than 5 (Lipinski et al., 1997; Naglah et al., 2020). The predicted drug analysis result is shown (Table II). All of the active ingredients in *U. lobata* satisfy Lipinski's rule of five in terms of molecular weight, torsion, number of hydrogen bond acceptors and donors, and lipophilicity.

Lipinski's rule of five is the most frequently applied rule used in drug-likeness analysis. The rule has four parameters with different criteria. A

compound with a molecular weight ≤ 500 g/mol has strong permeability to the blood and intestinal barriers, which lengthens the time it takes for it to pass through the lipid bilayer. The profile of hydrogen bond donor ≤ 5 and hydrogen bond acceptor ≤ 10 indicates that the compound can be effectively absorbed. The value exceeding the criterion indicates that the compound is soluble in a polar solvent such as water through hydrogen bonds (Pollastri, 2020). The logP parameter is related to the lipophilicity or hydrophobicity of a compound. Because the drug molecule is retained for a long time in the lipid bilayer membrane and is widely distributed in the body, a chemical with a logP value >5 causes severe toxicity. In contrast, a chemical that has a negative logP value is hydrophilic and may be difficult for the molecule to be absorbed. If a compound has a logP value of 5 and is not negative, it is said to have a high absorption and permeation profile since it can cross the hydrophilic outer layer of the membrane and enter the hydrophobic lipid bilayer (Rahmania et al., 2015).

The skin permeation of a molecule is expressed as a skin permeability coefficient (Kp), which is determined by molecular size and lipophilicity (Table III). If the log Kp value is negative, it indicates that the molecule is less skin permeable, and if it is less than -2.5 cm/h, it means that the molecule can hardly penetrate the skin (Testa and Kraemer, 2007). Mangiferin, in this instance, had a low skin permeant value and might not be skin absorbable. This property is crucial when developing certain drug delivery systems, especially if the drug is intended to be transdermal.

The pharmacokinetic profile of a molecule as a good drug candidate would also be dependent on its passive gastrointestinal absorption and bloodbrain barrier (BBB) permeation. Except for chrysoeriol, five of the active substances were shown to have low BBB penetration and gastrointestinal absorption, but chrysoeriol had high intestinal absorption. The likelihood that a substance will be actively transported through a biological membrane depends on whether it can act as a substrate for the permeability glycoprotein (Pgp), a crucial member of the ATP-binding cassette transporters (ABC-transporters). ABC transporters are present in the gastrointestinal wall as well as the brain (Lipinski et al., 1997). The major role of P-gp is to protect the central nervous system (CNS) from xenobiotics (Pires et al., 2015). The interaction of chemicals with cytochromes P450 (CYP), in addition to P-gp, is crucial.

Table I. Active compounds in *U. lobata* leaf extracts

| No | Active compounds | Molecule weight | Aqueous extract | Ethanolic extract |
|----|------------------|-----------------|-----------------|-------------------|
| 1 | Stigmasterol | 413 | (++) | (+++) |
| 2 | B-Sitosterol | 415 | (+) | (+) |
| 3 | Mangiferin | 423 | (+) | (++) |
| 4 | Quercetine | 303 | (-) | (-) |
| 5 | Kaempferol | 286 | (-) | (-) |
| 6 | Hypolaetin | 302 | (-) | (-) |
| 7 | Gossypetin | 318 | (+++) | (+) |
| 8 | Luteolin | 286 | (-) | (-) |
| 9 | Apigenin | 270 | (-) | (-) |
| 10 | Chrysoeriol | 300 | (+) | (+) |

Note: (-): negative; (+): weak; (++): moderate; (+++); strong

Table II. Physichochemical properties of active compounds in *U. lobata* leaf extracts

| | Lipinski's | | | | | | Denia |
|------------------|------------|--|-----|------------------------|-------------------|-----|-----------|
| Active compounds | MW | MW Log P Torsion HBA HBD No. of rule V | | No. of rule Violations | Drug- likeness | | |
| | ≤ 500 | ≤ 5 | ≤ 5 | ≤ 10 | ≤ 5 | ≤ 2 | likeliess |
| Stigmasterol | 412.69 | 8.56 | 5 | 1 | 1 | 0 | yes |
| β-Sitosterol | 414.71 | 9.34 | 6 | 1 | 1 | 0 | yes |
| Mangiferin | 422.34 | -0.37 | 2 | 11 | 8 | 0 | yes |
| Gossypetin | 318.24 | 1.81 | 1 | 8 | 6 | 0 | yes |
| Chrysoeriol | 300.26 | 3.10 | 2 | 6 | 3 | 0 | yes |

MW=Molecular weight; LogP=Lipophylicity; Torsi=rotatable bonds; HBA=H-bond acceptors; HBD=H-bond donors;

Table III. Pharmacokinetic properties of active compounds in *U. lobata* leaf extracts

| A -4: | Absorption | | Distribution | | Metabolism | | Excretion | Toxicity |
|------------------|------------|------------|--------------|----------|------------|-----------|-----------------|----------------|
| Active compounds | Intestinal | Skin | BBB | P-gp | CYP2D6 | CYP3A4 | Cltot | IDEO (mol/lrg) |
| compounds | absorption | permeation | permeant | Substrat | inhibitor | inhibitor | (log ml/min/kg) | LD50 (mol/kg) |
| Stigmasterol | Low | -2.74 | No | No | No | No | 0.618 | 2.540 |
| β-Sitosterol | Low | -2.20 | No | No | No | No | 0.628 | 2.552 |
| Mangiferin | Low | -9.14 | No | No | No | No | 0.347 | 2.396 |
| Gossypetin | Low | -6.96 | No | No | Yes | Yes | 0.304 | 2.527 |
| Chrysoeriol | High | -5.93 | No | No | Yes | Yes | 0.597 | 2.337 |

BBB= Blood Brain Barrier; P-gp= Permeability glycoprotein; Cltot=Clearance total

This superfamily of isoenzymes is a key player in drug biotransformation (Monatari and Ecker, 2015). CYP and P-gp could process small molecules synergistically to enhance the protection of tissues and organs (Szakács *et al.*, 2008).

Cytochrome P450 is an enzyme mostly found in the liver and functions in the detoxification and oxidation of drugs and other xenobiotics, as well as the body's removal of these substances. Although cytochrome P450 primarily renders these drugs inactive, it is also known that some pharmaceuticals may be activated by this enzyme, and as a result, this enzyme frequently participates

in the biotransformation of drugs. It is important to assess a drug candidate's capacity to act as a substrate for cytochrome P450, as well as P2D6 cytochrome (CYP2D6) and P3A4 cytochrome (CYP3A4), which are also the two main isoforms responsible for drug metabolism (Testa and Kraemer, 2007). From Table 3, it can be observed that almost all the active substances do not affect the CYP2D6 and CYP3A4 enzymes, apart from gossypetin and chrysoeriol. Two chemicals have the potential to increase drug levels in the blood and cause adverse drug reactions in addition to inhibiting the enzyme that metabolizes drugs. This

suggests that the P450 enzyme would metabolize the majority of the drugs tested in this study. Because β -sitosterol has a better overall clearance rate than other substances, it has a decreased risk of accumulation and toxicity. However, biological activity must be taken into account. Based on the LD-50 value, both mangiferin and chrysoeriol have higher toxicities than other compounds; hence, their safety needs to be evaluated. Most of the substances have a low value of intestinal absorption, except chrysoeriol, but because they are not P-gp substrates, they cannot cross the BBB. Chrysoeriol and gossypetin both inhibit CYP2D6 and CYP3A4, but not other drugs.

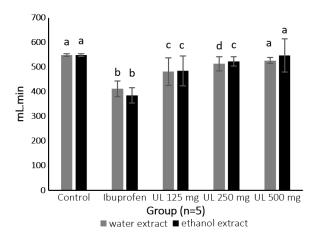


Figure 2. AUC of edema paw administrated U. lobata leaf extract; a,b,c etc. indicated the differences of potency (p<0.05, LSD test)

Anti-inflammatory effect of Urena lobata leaf extracts

The results of the anti-inflammatory activity of *U. lobata leaf extracts* as determined by the area under the curve (AUC) (Figure 2). At concentrations of 125, 250, and 500 mg/kg BW, aqueous extract of *U. lobata* significantly (p < 0.05) reduced edema paw by 12, 6, and 4%, respectively, compared to the control group. Similarly, the inhibition percentages of the ethanolic extract from the *U. lobata* were 12, 5 (p < 0.05), and 0.2% (p > 0.05), respectively, compared to the control group. Because aqueous and ethanolic extracts of *U. lobata* leaves contain β-sitosterol, mangiferin, and stigmasterol, they were able to reduce edema paw, even though their activity was less than that of ibuprofen. Mangiferin is a well-known natural polyphenol derived from plants that has several pharmacological actions, including those that are anti-inflammatory, analgesic, antibacterial,

antiviral, anti-diabetic, and antipyretic (Du *et al.*, 2018; Purnomo *et al.*, 2015). β -Sitosterol, a ubiquitious phytosterol in plants, is also found in *U. lobata* extracts (Purnomo *et al.*, 2018).

Both *in vivo* and *in vitro* studies have demonstrated that phytosterols have immunomodulatory properties; as a result, they may be beneficial for both human and animal health (Bouic and Lamprecht, 1999; Prieto *et al.*, 2006). Mangiferin and β -sitosterol can reduce COX-2 levels induced by lipopolysaccharides (Bhatia *et al.*, 2008; Lee *et al.*, 2012). The synthesis of prostacyclin (PGI2) and prostaglandins (PGE2) is reduced when COX-2 levels are reduced, resulting in the suppression of the inflammatory response (Bhatia *et al.*, 2008).

In vivo studies have shown mangiferin and βsitosterol to have COX-2 inhibitory activity (Bhatia et al., 2008; Lee et al., 2012). COX-2 plays a significant role in inflammatory pathogenesis, inflammatory agents such as bacterial endotoxins and cytokines may cause COX-2 to be secreted. It is involved in the release of prostanoids during inflammation, making it a potential molecular target for analgesics and anti-inflammatory medications (Ricciotti and Fitzgerald, 2011). Based on an in silico investigation, previous research also suggested that mangiferin, stigmasterol, and β-sitosterol inhibited COX-2 activity (Wahyuningsih et al., 2022). Furthermore, mangiferin and β-sitosterol have inhibitory activity on COX-2. The expression of nuclear factor kappa B (NF-κB) and tumor necrotic factor (TNF- α) is known to be inhibited by mangiferin and β-sitosterol (Loizou et al., 2010; Lee et al., 2012; Saha et al., 2016; Sun et al., 2020). TNFα is a pro-inflammatory cytokine with numerous physiological and pathological functions. TNF-α activates NF-κB, which is required for the expression of pro-inflammatory cytokines, the differentiation of innate immune cells, and the differentiation of inflammatory T-cells. NF-κB and TNF- α inhibition actions, which also reduce inflammation, aid in the healing process (Lawrence, 2009; Pozniak et al., 2014; Liu et al., 2017). Both the ethanolic and aqueous extracts of U. lobata leaf extract had relatively similar efficacy in reducing the levels of paw edema. A 500 mg/kg BW dose of aqueous and ethanolic extracts of *U. lobata*, however, was unable to reduce paw edema. Low receptor sensitivity is caused by active drug dose and duration increases (Dar et al., 2005). Additionally, the extract's anti-inflammatory efficacy will decrease with an increase in dose. The administration of *U. lobata* leaf extract had potency

as an anti-inflammatory agent. However, its efficacy is lower than ibuprofen as a reference drug. The herbs contain many active compounds that can interact with one another. This interaction may have perhaps resulted in an antagonist effect, therefore reducing its activity (Goodman and Gilman, 2006).

Analgesic effect of Urena lobata leaf extracts

When compared to the control group, the administration of *U. lobata* aqueous extract at 125, 250, and 500 mg/kg BW significantly (p<0.05) increased the AUC of pain threshold by about 40, 70, and 100%, respectively (Figure 3). Meanwhile, ethanolic extract from *U. lobata* increased significantly (p<0.05) by 20, 50, and 50%, respectively. Except for the aqueous extract of *U*. lobata at a dose of 500 mg/kg BW, the activity was lower than ibuprofen. This observation might be a result of some active ingredients in *U. lobata*, which can act by inhibiting the release of pain mediators such as prostaglandin, bradykinin, and histamine (Goodman and Gilman, 2006; Ricciotti and Fitzgerald, 2011). β-sitosterol and mangiferin are active compounds in *U. lobata* leaf extract having activity as an analgesic.

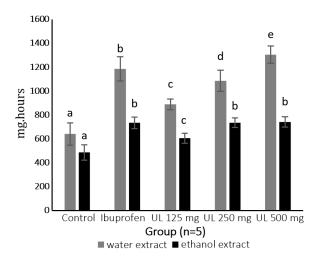


Figure 3. AUC of pain threshold administrated *U. lobata* leaf extract; a,b,c etc. indicated the differences of potency (p<0.05, LSD test).

The study of the analgesic activity of mangiferin showed anti-nociceptive responses in both peripheral and central areas (Dar *et al.*, 2005). The peripherally active sensory afferent nerves are the source of nociceptive responses, neuropathic pain, and anti-inflammatory responses. This shows that the inhibition of several neuro-receptors, such

as opioids, alpha-adrenergic, cholinergic, and cannabinoid receptors, may represent possible targets for drugs (Sawynok, 2003; Imran et al., 2017). The involvement of mangiferin and βsitosterol in the opioid receptor cannot be ignored (Dar et al., 2005; Dighe et al., 2016). β-sitosterol showed central anti-nociceptive action, therefore exerting its effect through the central opioid receptor or promoting the release of endogenous opioid peptides (Dighe et al., 2016). Mangiferin and β-sitosterol can lower COX-2 levels, which in turn prevents the formation of both prostaglandins (PGE2) and prostacyclin (PGI2), a pain-mediating compound (Bhatia et al., 2008; Lee et al., 2012). Some of the aforementioned inflammatory mediators can activate pain receptors or nociceptors, which results in pain. Histamine and PGE2 release stimulate nociceptors (nociceptive afferent fibers), which are responsible for transmitting pain signals to the brain (Dar et al., 2005). As demonstrated in this study, stigmasterol, β-sitosterol, and mangiferin can inhibit COX-2 activity by molecular docking, therefore serving as potential anti-nociceptive.

An aqueous extract of *U. lobata* has a stronger analgesic effect than its ethanolic extract. This may indicate that the *U. lobata* active ingredients have low solubility and absorption in ethanol, which would reduce their bioactivity as analgesics. The formation of complex compounds as well as changes in active compound conformation would further reduce active substance solubility and therefore decrease its ability to prevent pain mediator release (Stevens et al., 2010). With a dose of 125 mg/kg BW, the ethanol extract of *U. lobata* exhibits the least analgesic efficacy. It is brought on by modest doses of herbs' inability to prevent the release of pain mediators and nociceptor activation.

CONCLUSION

The findings of this study reveal that the leaf extracts from *Urena lobata* have anti-inflammatory and analgesic properties. The range established by Lipinski's rule of five for drug-likeness analysis was applied to all of the phytoconstituents, and three constituents were expected to have better ADME-T characteristics.

ACKNOWLEDGEMENT

The author express their profound gratitude to the University of Islam Malang for funding this research.

AUTHORS' DECLARATION

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

REFERENCES

- Adewale AO, David AA, and Abiodun OO. 2007. Studies on anti-microbial, antioxidant and phytochemical analysis of Urena lobate leaf extract. *J Phys Nat Sci*, 1, 12–20.
- Babu SS, Madhuri BD, and Reddy SD. 2016. Anxiolytic Antidepressant and Anti-Inflammatory Activity of Ethanolic Extract of Urena Lobata Leaf. International Journal of Pharma Research and Health Sciences, 4(4), 1284–1290.
 - https://doi.org/10.21276/ijprhs.2016.04.0
- Bhatia HS, Candelario-jalil E, Pinheiro AC, Oliveira D, Olajide OA, Martínez-sánchez G, and Fiebich BL. 2008. Mangiferin inhibits cyclooxygenase-2 expression and prostaglandin E 2 production in activated rat microglial cells. *Archives of Biochemistry and Biophysics*, 477(2), 253–258. https://doi.org/10.1016/j.abb.2008.06.017
- Bouic PJD, and Lamprecht JH. 1999. Plant sterols and sterolins: A review of their immune-modulating properties. *Alternative Medicine Review*, *4*(3), 170–177.
- Carugo O and Pongor S. 2001. A normalized rootmean squared distance for comparing three dimensional structures. *Protein Science*. 10, 1470-1473
- Dar A, Faizi S, Naqvi S, Roome T, Sadia ZR, Ali M, Firdous S, and Moin ST. 2005. Analgesic and Antioxidant Activity of Mangiferin and Its Derivatives: the Structure Activity Relationship. *Biol. Pharm. Bull.*, *28*(4), 596–600.
- Dighe SB, Kuchekar BS, and Wankhede SB. 2016. Analgesic and anti- inflammatory activity of β-sitosterol isolated from leaves of Oxalis corniculata. *International Journal of Pharmacological Research*, 6(3), 109–113. https://doi.org/10.7439/ijpr
- Du S, Liu H, Lei T, Xie X, Wang H, He X, Tong R, and Wang Y. 2018. Mangiferin: An effective therapeutic agent against several disorders (Review). *Molecular Medicine Reports*, 18, 4775–4786.
- https://doi.org/10.3892/mmr.2018.9529 Goodman LS, and Gilman A. 2006. *Goodman &*

- Gilman's the pharmacological basis of therapeutics. McGraw-Hill.
- Imran M, Arshad MS, Butt MS, Kwon J, Arshad MU, and Sultan MT. 2017. Mangiferin: a natural miracle bioactive compound against lifestyle related disorders. *Lipids in Health and Disease*, 16(84), 1–17. https://doi.org/10.1186/s12944-017-0449-y
- Jain, AN. and Nicholls, A. 2008. Recommendations for evaluation of computational methods. Journal of computer aided molecular design, 22(3), 133-139. https://doi.org/10.1007/s10822-008-9196-5.
- Lawrence T. 2009. The Nuclear Factor NF-κB Pathway in Inflammation. *Cold Spring Harbor Perspectives in Biology*, 1(6), 1–10. https://doi.org/ 10.1101/cshperspect.a001651
- Lee I, Kim E, and Kim D. 2012. Inhibitory Effect of β-Sitosterol on TNBS-Induced Colitis in Mice. *Planta Medica*, 78(9), 896–898.
- Lipinski, C.A., Lombardo, F., Dominy, B.W. and Feeney, P.J., 1997. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. Advanced drug delivery reviews, 23(1-3), pp.3-25. https://doi.org/10.1016/S0169-409X(96)00423-1
- Liu T, Zhang L, Joo D, and Sun S. 2017. NF-κB signaling in inflammation. *Signal Transduction and Targeted Therapy*, 2, 1–9. https://doi.org/10.1038/sigtrans.2017.23.
- Loizou S, Lekakis I, Chrousos GP, and Moutsatsou P. 2010. β-Sitosterol exhibits anti-inflammatory activity in human aortic endothelial cells. *Molecular Nutrition & Food Research*, 54(4), 551–558. https://doi.org/10.1002/mnfr.200900012
- Montanari, F. and Ecker, G.F., 2015. Prediction of drug–ABC-transporter interaction—Recent advances and future challenges. *Advanced drug delivery reviews*, 86, pp.17-26. https://doi.org/10.1016/j.addr.2015.03.00
- Naglah, A.M., Askar, A.A., Hassan, A.S., Khatab, T.K., Al-Omar, M.A. and Bhat, M.A., 2020. Biological evaluation and molecular docking with in silico physicochemical, pharmacokinetic and toxicity prediction of pyrazolo [1, 5-a]

- pyrimidines. *Molecules*, *25*(6), p.1431. https://doi.org/10.3390/molecules250614 31
- Nauli, T. 2014. The Identification of active site of cellulase Aspergillus niger with ligand docking. *Indonesian Journal of Applied Chemistry*,16(2), 94-100.
- Pires, D.E., Blundell, T.L. and Ascher, D.B., 2015. pkCSM: predicting small-molecule pharmacokinetic and toxicity properties using graph-based signatures. *Journal of medicinal chemistry*, *58*(9), pp.4066-4072. https://doi.org/10.1021/acs.jmedchem.5b 00104
- Pollastri, M.P., 2010. Overview on the Rule of Five. *Current protocols in pharmacology*, 49(1), pp.9-12. https://doi.org/10.1002/0471141755.ph0 912s49
- Pozniak PD, White MK, and Khalili K. 2014. TNF- α/NF - κB Signaling in the CNS: Possible Connection to EPHB2. *J Neuroimmune Pharmacol*, 9(2), 133–141. https://doi.org/10.1007/s11481-013-9517- x
- Price W. 2006. *Patofisiologi Vol 2*. Buku Kedokteran EGC.
- Prieto JM, Recio MC, and Giner RM. 2006. Antiinflammatory activity of beta-sitosterol in a model of oxazolone-induced contact delayed type hypersensitivity. *Boletin Latino americano y Del Caribe de Plantas Medicinales y Aromaticas*, 5(3), 57–62.
- Purnomo Y, Soeatmadji DW, Sumitro SB and Widodo MA. 2015. Anti hyperglycemic effect of *Urena lobata* leaf extract by inhibition of dipeptidyl peptidase IV (DPP-IV) on diabetic rats. *International Journal of Pharmacognosy and Phytochemical Research*. 2015; 7:1073–1079.
- Purnomo Y, Soeatmadji DW, Sumitro SB and Widodo MA. 2018. Inhibitory activity of Urena lobata leaf extract on dipeptidyl peptidase-4 (DPP-4): is it different in vitro and in vivo?. Medicinal Plants International Journal of Phytomedicines and Related Industries, 10(2), 99–105. https://doi.org/10.5958/0975-6892.2018.00016.3
- Purnomo Y, 2021. Makdasari J, Fatahillah FI. Inhibitory activity of Urena lobata leaf extract on alpha-amylase and alpha glucosidase: *in vitro* and *in silico* approach. *Journal Basic Clinical Physiology and Pharmacology*. 2021;

- 32(4):889-894
- Rachmania, R.A. and dan Oetari, S., 2013. In silico analysis of diterpenoid lactone compounds of bitter herbs (Andrographis paniculata Nees) on alpha glucosidase receptor as antidiabetic type II agents. *Pharmacy*, 12(2), pp.210-222.
- Ricciotti E, and Fitzgerald GA. 2011. Prostaglandins and Inflammation. *Arteriosclerosis Thrombosis and Vascular Biology*, *31*(5), 986– 1000.
 - https://doi.org/10.1161/ATVBAHA.110.207 449
- Rivera DG, Balmaseda IH, León AÁ, Hernández BC, Montiel LM, Garrido GG, Cuzzocrea S, and Hernández RD. 2006. Anti-allergic properties of Mangifera indica L . extract (Vimang) and contribution of its glucosylxanthone mangiferin. *Journal of Pharmacy and Pharmacology*, 58(3), 385–392. https://doi.org/10.1211/jpp.58.3.0014.
- Saha S, Sadhukhan P, and Sil PC. 2016. Review Article Mangiferin: A xanthonoid with multipotent anti-inflammatory potential. *BioFactors*, 42(5), 1–16. https://doi.org/10.1002/biof.1292
- Sawynok J. 2003. Topical and Peripherally Acting Analgesics. *Pharmacological Reviews*, *55*(1), 1–20. https://doi.org/10.1124/pr.55.1.1.1
- Sen S, Chakraboty R, De B, Ganesh T, Raghavendra HG, and Debnath S. 2010. Analgesic and Anti-Inflammatory Herbs: a Potential Source of Modern Medicine. *International Journal of Pharmaceutical Sciences and Research*, 1(11), 32–44.
 - http://dx.doi.org/10.13040/IJPSR.0975-8232.1(11).32-44
- Stevens MM, Honerkamp-smith AR, and Keller SL. 2010. Solubility limits of cholesterol, lanosterol, ergosterol, stigmasterol, and b-sitosterol in electroformed lipid vesicles. *Soft Matter*, 6(23), 5819–6014. https://doi.org/10.1039/c0sm00373e
- Sun Y, Gao L, Hou W, and Wu J. 2020. β-Sitosterol Alleviates Inflammatory Response via Inhibiting the Activation of ERK/p38 and NF-κB Pathways in LPS-Exposed BV2 Cells. *BioMed Research International*, 1–10. https://doi.org/10.1155/2020/7532306.
- Szakács, G., Váradi, A., Özvegy-Laczka, C. and Sarkadi, B., 2008. The role of ABC transporters in drug absorption, distribution, metabolism, excretion and toxicity (ADME-Tox). *Drug Discovery*

Today, *13*(9-10), pp.379-393. https://doi.org/10.1016/j.drudis.2007.12.0 10

Testa, B. and Krämer, S.D., 2007. The biochemistry of drug metabolism–an introduction: Part 2. Redox reactions and their enzymes. *Chemistry & biodiversity*, 4(3), pp.257-

405. https://doi.org/10.1002/cbdv.20079 0032

Wahyuningsih D, Purnomo Y, Tilaqza A., 2022. In Silico study of Pulutan (Urena lobata) leaf

extract as anti inflammation and their ADME prediction. Journal of Tropical Pharmacy and Chemistry, 6(1), pp.30-37. https://doi.org/10.25026/jtpc.v6i1.323

Zambre, A.P., Ganure, A.L., Shinde, D.B., and Kulkarni, V.M., 2007. Perspective assessment of COX-1 and COX-2 selectivity of nonsteroidal anti-inflammatory drugs from clinical practice: use of genetic function approximation, *J. Chem. Inf. Model.*, 47, 635-643. https://doi.org/10.1021/ci6004367