MORPHOLOGICAL AND MOLECULAR CHARACTERS OF *Mimegralla* spp. (DIPTERA: MICROPEZIDAE) ON ZINGIBERACEAE IN CENTRAL JAVA

KARAKTER MORFOLOGI DAN MOLEKULER Mimegralla spp. (DIPTERA: MICROPEZIDAE) PADA PERTANAMAN ZINGIBERACEAE DI JAWA TENGAH

Rahma Widyastuti^{1)*}, Suputa²⁾, & Nugroho Susetya Putra²⁾

¹⁾Center for Research and Development of Medicinal Plants and Traditional Medicine Jln. Raya Lawu No. 11, Tawangmangu, Karanganyar, Central Java 57792

²⁾Department of Plant Pest and Disease, Faculty of Agriculture, Universitas Gadjah Mada Jln. Flora 1, Bulaksumur, Sleman, Yogyakarta 55281

*Corresponding author. E-mail: rahma.marwasti@gmail.com

ABSTRACT

Rhizome fly, *Mimegralla* sp. (Diptera: Micropezidae) is a major pest on Zingiberaceae plants. Some of those fly species have been reported attacking some of Zingiberaceae plants, but in Indonesia, only one species, *Mimegralla coeruleifrons* has been reported as being a pest. Since Indonesia has many species of Zingiberaceae plants, it may raise a prediction that more than one species of Mimegralla was found on this plants. Therefore, a taxonomic research on the species of rhizome flies to find the species other than *M. coerulifrons* on Zingiberaceae plants is urgently required. This study was conducted by using hand-picking method on Mimegralla adult inhabiting Zingiberaceae plants (ginger, turmeric, javanese ginger, and aromatic ginger), and was then identified by using morphological characters and through molecular technique by using mtCO1 gene. The results showed that *M. albimana* and *M. coeruleifrons* found at four zingiberaceae plants were the member of Mimegralla. As a conclusion, these two species have high values of phylogenic relationship (88%) and bootstrap (92).

Keywords: Mimegralla spp., molecular, morphology

INTISARI

Lalat rimpang (Mimegralla spp.) adalah hama utama pada beberapa tanaman anggota Family Zingiberaceae. Beberapa spesies lalat rimpang dilaporkan menyerang tanaman Zingiberaceae, di antaranya jahe, kunyit, temulawak, dan kencur. Di Indonesia hanya satu spesies yang pernah dilaporkan menyerang tanaman jahe yaitu Mimegralla coeruleifrons. Sementara itu, jenis tanaman Zingiberaceae yang ditanam di Indonesia sangat beragam, sehingga muncul dugaan bahwa Mimegralla yang menyerang tanaman Zingiberaceae di Indonesia tidak hanya M. coeruleifrons saja. Oleh karena itu, penelitian untuk mengidentifikasi spesies-spesies lalat rimpang yang menyerang tanaman Zingiberaceae di Indonesia perlu dilakukan. Penelitian ini dilakukan menggunakan metode pemungutan/penangkapan (hand-picking) imago Mimegralla pada tanaman keempat tanaman Zingiberaceae. Imago yang diperoleh diidentifikasi secara morfologi dan molekuler menggunakan gen mtCO1. Hasil penelitian ini menunjukkan bahwa M. albimana dan M. coeruleifrons yang ditemukan pada pertanaman Zingiberaceae merupakan satu genus yang sama yaitu Mimegralla. Hasil identifikasi molekuler menunjukkan bahwa kedua lalat rimpang tersebut mempunyai derajat kekerabatan dengan nilai homologi sebesar 88%, dan nilai bootstrap pada pohon filogeni sebesar 92.

Kata kunci: Mimegralla spp., molekuler, morfologi

INTRODUCTION

Zingiberaceae plants, including genus Curcuma, Kaempferia, Hedychium, Amomum, Zingiber, Alpinia, Elettaria, and Costus (Joy *et al.*, 1998), are the main materials in producing herbal medicines, which is containing volatile oils and oleoresins as tonic and stimulant for human body. Rhizome fly is noted as a major pest in ginger plants (Steyskal, 1964), turmeric (Nair, 1980), and aromatic ginger (Balfas *et al.*, 2000) in several countries, including Indonesia.

Two species of rhizome fly are reported attacking Zingiberacea in India (Maxwell-Lefroy & Howlett,

1909), those are *Mimegrallla coeruleifrons* Macq. (Ghorpade *et al.*, 1983) and *M. albimana* Doleschall (Ghorpade *et al.*, 1988). But only one species, *M. coeruleifrons* Macq., was reported attacking Zingiberaceae in Indonesia (Balfas, 2002). However, the diverse species of Zingiberaceae planted in Indonesia raises asumption that the number of Mimegralla attacking rhizomes might be more than one species. Therefore, a thorough study should be done to clarify this issue. This study was aimed to know the species of Mimegralla attacking Zingiberaceae in Central Java by using morphological and molecular characters. In addition, the molecular identification by using

mtCO1 characters on rhizome fly (*Mimegralla* spp.) has never been done, and even the information of base nucleotide was never found in NCBI GenBank. So identification using mtCO1 need to be done to understand the phylogenic relationship.

MATERIALS AND METHODS

Sampling and Preservation

Mimegralla spp. adults were collected by handpicking from ginger (Zingiber officinale), turmeric (Curcuma domestica), Javanese ginger (temulawak, Curcuma xanthorrhiza), and aromatic ginger (kencur, Kaempferia galanga) plants in Karanganyar District (latitude of 7°37'53,674", longitude 110°56'58,957" and altitude of 204.7 m above sea level) and Purworejo District (latitude 7º42'15,660", longitude 110°1'47,537" LE and latitude of 81.5 m above the sea level). Sampling was conducted in area, where symptomatic of rhizome fly, i.e. rich of organic material rot (Hennig, 1935) and wilt (Karmawati et al., 1990) were found. Sampling were conducted in October 2016, when the rainy season started and Zingiberaceae thrives at this season. Heavy rainfall is the optimum condition of the microorganisms to grow well.

Adult of rhizome flies were used for morphological and molecular identifications, and insect collection as well. Adults used for morphology identification were pinned using insect needles no. 00 and put into 96% alcohol for molecular identification.

Identification

Morphological and molecular identification were conducted at Laboratory of Basic Entomology and Laboratory of Virology, Department of Plant Pest and Disease, Faculty of Agriculture, Universitas Gadjah Mada, Yogyakarta.

Morphological identification of fly was done by using Leica MZ16 and Leica KL1500 LCD Microscopes and Optilab advance on the whole part of body to describe shape, size, color, and genitalia parts as specific characters of species. Magnification of microscopes showed in Table 1.

DNA extraction was done by using Genomic Mini Kit from Geneaid, and the extraction product was then amplified by PCR using LCO1490 (GGT CAA CAA ATC ATA AAG ATA TTG G) as forward primer and HCO2198 (TAA ACT TCA GGG TGA CCA AAA AAT CA) as reverse primer (Folmer et al., 1994). The kit used in PCR reaction was Go Taq Green Master Mix kit: Go Tag Green Master Mix 12.5 μ L; primer forward 1 μ L; reverse primer 1 μ L; DNA template 2 µL; and Nuclease Free Water 8.5 μL. Stages of PCR cycles were shown in Table 2. The results of subsequent DNA amplification were visualized by agarose gel electrophoresis (SIGMA) and observed using BIO-RAD UV transluminator 2000. Sequencing was carried out by sending samples to 1st Base DNA Sequencing (Selangor, Malaysia). Finally, to analyze the degree of neighborhood joining values, Bioedit software was used, and followed by Mega 5.2. and BLAST software. The Bioedit was

Component identification	Magnification				
	Mimegralla coeruleifrons	Mimegralla albimana			
All body	11.4 ×	20.0 ×			
Caput	51.2 ×	64.0 ×			
Thorax	$40.0 \times$	51.2 ×			
Abdomen	25.6 ×	$40.0 \times$			
Wings	25.6 ×	32.0 ×			
Antenna	128.0 ×	128.0 ×			
Copulatory fork	$100.8 \times$	$100.8 \times$			
Ovipositor	32.0 ×	80.0 imes			

Table	1 N	Aaoni	ification	for	mornhol	logy	ident	ification
raute	T • T	viagin	incunon	101	morphor	USY	ruom	incurion

Table	2.	Р	CR	stages	cyc	le
-------	----	---	----	--------	-----	----

No.	Reaction	Temperature	Time	Cycle
1	Predenaturasi	95°C	2 minutes	
2	Denaturasi	95°C	30 seconds	
3	Annealing	55°C	30 seconds	- 30 cycles
4	Extention	72°C	1 minute	
5	Final extention	72°C	5 minutes	
6	Delay	4°C	5 minutes	

used to make base paired alignment of forward and reverse nucleotide bases, and to get the percent of clustal consensus. We used MEGA 5.2. for making phylogeny tree and used BLAST for getting homology percentage.

RESULTS AND DISCUSSION

In this study 78 samples of suspected Mimegralla was found in Zingiberaceae plants (ginger, turmeric, javanese ginger, and aromatic ginger). The result of morphological identification showed that the samples was consisted of two different Mimegralla species, i.e. *Mimegralla albimana* and *M. coeruleifrons* (Figure 1). To complete the observation on the differensiation between these two species, three individual samples of each species were selected randomly, and were examined by using molecular technique.

Morphology Identification

Stilt-legged fly (*Mimegralla* spp.) is a member of family Micropezidae (Diptera: Acalyptratae), and there are 583 spesies of Micropezidae reported, which are divided into 52 genera and 5 subfamilies (Pape *et al.*, 1758). Most of Micropezidae species were found in tropic and subtropic areas. This fly has unique characteristics by their long and slender feet.

Aczel (1959) mentioned that a specific character of Mimegralla is the occurence of ocellar plate on the posterior of upper superior orbital or very close to the fore ocellus placed in a line. Frontal stripe are more than a half of total width of frons. Upper anterior orbital stand in tomentosa frontal or form a boundary line between the frontal stripe and genovertical plate. Postvertical and genal bristle always absen.

- 1. Forceps in apex of copulatory fork tapered at the ends with intersect bristles (in males). Shape of ovipositor oval with membraneus region at the end the shorter (Figure 2). Body color is brownish red and yellowish *albimana*
- 2. Forceps in apex of copulatory fork show widened with a short bristle does not intersect (in males). Shape of ovipositor conical with an elongated tip at membraneus region (Figure 3). Overall body color is dark blue and black *coeruleifrons*

M. albimana and *M. coeruleifrons* could be distinguished by their color. *M. albimana* has brownish red and yellow on its body. Facet are maroon, antenna are reddish yellow, abdomen black and brownish yellow, legs bright yellow with black stripes on coxa, more dark to the apex. Whereas *M. coeruleifrons* has blackish-dark-blue body. Facet are dark blue and metallic when exposed to the light, antennae are dark brown, abdomen is black and gray, and legs are brownish with black stripes on coxa. Hennig (1935) also reported, *M. albimana* legs are continuosly dark whereas *M. coeruleifrons* legs are dark.

However, when the body of *M. albimana* and *M. coeruleifrons* were observed closer by using higher magnification, some differences were found. These differences were observed on preabdomen, ovipositor, wings, and copulatory fork. Body of *M. albimana* was stout and looks more convex than long, length of wings were 3.5 to 3.7 times than width, and lenght of body was 4.3 to 4.5 times than width (Figure 1). Whereas *M. coeruleifrons* was more slender and elongated and wing length was 3.8 to 4.5 times than width. Body lenght 4.6 to 4.9 times than width.



Figure 1. Female adult of *Mimegralla albimana* (a); adult of *Mimegralla coeruleifrons* (b): frons (f), tergite 1–3 (t), apex of wing (ap), ovipositor (o), legs (l)



Figure 2. Wing (a), head (b), copulatory fork (c), and ovipositor (d) of Mimegralla albimana



Figure 3. Wing (a), head (b), copulatory fork (c), and ovipositor (d) of Mimegralla coeruleifrons

Molecular Identification

DNA amplification on adult was done by using cytochrome c oxidase subunit 1 in mitochondrial (mtCO1) part and was performed by universal primers LCO1490 and HCO2198, which are mostly used for invertebrates (Folmer *et al.*, 1994).

DNA amplification was visualized using BIO-RAD UV transluminator 2000, and a single band at 600 bp. This product then was sent to 1st Base DNA Sequencing at Selangor, Malaysia. The result of DNA sequencing of *M. albimana* showed, that it has 681 bp (base pairs) of DNA nucleotide bases, whereas *M. coeruleifrons* has 678 bp. These results were consistent with results of PCR visualization as described above. The nucleotide bases of *M. albimana* was compared with *M. coeruleifrons* and three other species (Table 3), the member of Micropezidae Family, to determine the value of homology between the first two species and the other three. The nucleotide bases of the three other species was taken from GenBank database as shown at Figure 4.

The nucleotide bases of five species were compared to determine the level of homology in percent. The closeness of phylogenic relationship in clustal consensus

	10	20	30	40	50	60	10	80
Mimegralla albimana	TAAAGATATT	GGAACTTTAT	ATTTTATATT	TEGAGETTEA	GCAGGAATAG	TECCEACTTE	TTTAAGAATT	TTAATTCGAG
Mimegralla coeruleifrons	AAGATATT	GGAACATTAT	ATTTCATATT	TGGAGCTTGA	GCGGGGGATAG	TAGGGACCTC	TTTAAGAATC	TTAATTCGAG
Hemichaeta scutellata		ACT CT AT	ATTTTATATT	CGGAGCTTGA	GCAGGAATAG	TAGGAACTTC	TTTAAGAATC	CTCATTCGAG
Taeniaptera trivittata		ACTITAT	ATTTTATATT	CGGAGCTTGA	GCAGGAATAG	TAGGAACTIC	TTTAAGAATT	CTAATCCGTG
Micropera sp. Clastal Consensas		ACATTAL	ATTTTATTT	IGGAACTIGA	GCAGGAATAG	TAGGAACTIC	CITAAGAATT	TTAATTCGAG
Children Couperses								
	90	100	110	120	130	140	150	160
M								
Mimegralia albimana Mimegralia coeraleifrons	CTGAATTAGG	TCACCCAGGA	GCTCTAATTG	GAGACGATCA	AATTTATAAT	GT AATTGT AA	CAGCICACGC	CTTTGTTATA
Hemichaeta scutellata	CTGAATTAGG	TCACCCAGGA	GCTTTAATTG	GTGACGACCA	GATITATAAT	GTAATTGTTA	CCGCTCATGC	ATTCATTATA
Taesiaptera trivittata	CTGAACTAGG	ACACCCAGGA	GCCCTAATTG	GAGATGATCA	AATTTACAAT	GTAATTGTAA	CTGCACATGC	ATTTATTATA
Micropera sp.	CAGAATTAGG	ACATCCAGGA	GCATTAATIG	GAGATGATCA	AATTTATAAT	GTAATTGTTA	CTGCACATGC	TITIGTAATA
Clustal Consensus								
	:70	:10	190	200	210	220	250	240
	rectrond	and the set		restreef.	1000100001		see been b	erester eff
Mimegralla albimana	ATTITITA	TAGTTATACC	TATCATAATT	GGIGGATITG	GAAATTGACT	AGTACCITTA	ATATTAGGTG	CICCAGATAT
Hemichaeta scutellata	ATTTTCTTTA	TAGTTATACC	TATTATAATC	GGTGGATTTG	GAAATIGATT	AGTICCITTA	ATACTAGGAG	CICCIGATAT
Taeniaptera trivittata	ATTTTTTTA	TAGTGATACC	TATTATAATT	COTCOCTTCC	GAAACTGATT	AGTICCATIA	ATGTTAGGAG	CCCCTGATAT
Micropera sp.	ATTTTTTTA	TAGTAATACC	TATTATAATT	GGAGGATITG	GAAACTGATT	AGTACCATTA	ATATTAGGTG	CTCCTGATAT
Clustal Consensus								
	250	260	270	290	290	300	310	320
Mimagralla albimana	1001111001	CONTINUT	AT AT A ACT	CTGACTTOTT	COTOCTOCTO	TTACTTATT	ATTACTANCE	IGANTICTIC
Mimegralla coeruleifrons	AGCATICCCT	CGAATAAATA	ATATAAGITT	TIGACITCIT	CCTCCCGCCC	TTACATTATT	ATTAGTAAGC	AGAATAGTAG
Hemichaeta scutellata	AGCTTTTCCA	CGAATAAATA	ATATAAGATT	TTGACTCCTT	CCACCAGCTT	TAACTITATT	ATTAGTAAGA	AGAATAGTAG
Taeniaptera trivittata	AGCATTICCA	CGAATAAATA	ATATAAGATT	TIGACITITA	CCGCCAGCTT	TAACCTTACT	ATTAGTAAGA	AGAATAGTAG
Micropera sp.	AGCATTICCT	CGAATAAATA	ATATAAGATT	TIGACITITA	CCTCCAGCAT	TAACTITATT	ATTAATAAGT	AGAATAATIG
Classal Coastasas								
	330	340	350	360	310	380	390	400
Mimerralla albimana	ANATGGAGC	TEGEACAGEA	TGAACAGTTT	ATCCCCCTCT	TICITCAGTA	ATTGCACATG	GAGGAGCTTC	TGTAGACTTA
Mimegralla coeruleifrons	AAAATGGAGC	CGGAACAGGA	TGAACGGTTT	ACCCTCCACT	CTCATCAGTA	ATTGCCCATG	GAGGAGCTTC	AGTAGATITA
Hemichaeta scutellata	AAAATGGAGC	TGGTACAGGA	TGAACTGTAT	ACCCACCGCT	AT CAT CGGT A	ATTGCTCATA	GAGGAGCATC	TGTAGATTTA
Taeniaptera trivittata	AAAACGGAGC	CGGTACTGGA	TGAACAGTAT	ACCCACCITI	ATCATCAGTA	ATCGCTCATG	GAGGTGCTTC	GGTTGATTTA
Clustal Consensus	*****		TOAACOOTAT	Accententi	ATCATCIACA	ATTOCICATA	GAGGAGCUIC	IGIIGAITIA
	410	420	400	440	410	460	470	410
Mimegralla albimana	GCAATTTTCT	CATTACATTT	AGCAGGTATC	TETTETATTT	TAGGAGCAGT	AAATTTTATT	ACTACTGTAA	TTAATATGCG
Mimegralla coeruleifrons	GCAATTTTCT	CATTACACCT	AGCTGGTATC	TETTCAATTC	TAGGAGCAGT	AAACTTTATT	ACAACTGTAA	TTAATATACG
Hemichaeta scutellata	GCTATTTTCT	CCCTTCATTI	AGCAGGAATC	TCTTCAATTC	TIGGIGCAGT	AAATTTTATT	ACTACAGTAA	TTAATATACG
Taentaptera trivittata Microneta un	GUATTITT	CACTICATU	AGCTGGAGTA	TCCTCTATTT	TAGGAGCAGT	AAATTTTATT	ACTACAGTAA	TTAATATACG
Clustal Consensus								
	400	500		120		[4]		100
Mimegralla albimana	ATCAACAGGA	ATCACTTTAG	ACCGTATACC	TTTATTTGTA	TGATCAGTAG	TTATTACAGE	ATTTTTACTC	TTACTITCIT
Mimegralla coeruleifrons	ATCAACAGGT	ATTACATTAG	ACCGAATACC	ATTATTIGTT	TGATCIGITG	TAATTACCGC	ATTTTTATTA	CITCICICIC
Taeniantera trivittata	ATCTACAGGA	ATTACTITAG	ACCGTATACC	TTTATTTGTT	TGATCAGTAG	TAATTACAGE	ATTTCTTTA	TIGTTATCTT
Micropera sp.	AACTACAGGA	ATTACTTTAG	ATCGAATACC	ATTATTTGTA	TGATCIGITG	TAATTACAGC	ATTTCTTTA	TTATTATCTT
Clustal Consensus								
Mimegralla albimana	TACCIGIATI	AGCAGGAGCA	ATTACTATAT	TATTAACAGA	CCGAAATCTT	AATACATCAT	TITITGACCC	TGCAGGAGGT
Mimegralia coeruleitrous Hemichaeta scntellata	TACCIGICCT	AGCAGGAGCA	ATTACTATAT	TATTAACAGA	CCGALATITA	AACACATCTT	TITTIGACCC	AGCIGGIGGI
Taeniaptera trivittata	TACCAGTATT	AGCAGGAGCA	ATTACTATAT	TATTAACTGA	TEGAAATETT	AATACATCAT	TITTTGATCC	AGCAGGAGGA
Micropera sp.	TACCAGITIT	AGCTGGAGCA	ATTACAATAT	TATTAACAGA	TCGAAATTTA	AATACTTCAT	TITTTGACCC	TGCAGGAGGA
Clustal Consensus								
	<i>e</i> ta	10	174	(11				
Minegralia albimana Minegralia cosmisifican	GGIGATCCTA	TTTTATACCA	ACALITATIT	IGATITITIG	6			
Hemichaeta scutellata	GGAGATCCTA	TITTATACCA	ACATITATIT	IGATITITIG	т			
Taeniaptera trivittata	GGAGATCCAA	TTTTATACCA	ACATTTATTT	TGATTTTTTG	G			
Micropera sp.	GGAGATCCAA	TITTATACCA	ACATTTATTT	IGATITITIG	C			
CIUSTAI COUSHUSUS								

Figure 4. Clustal consensus between *Mimegralla* spp. and three others species of Micropezidae Family from GenBank database

Species	Total of nucleotide base (bp)	Origin	Source
Mimegralla albimana	681	Karanganyar	This research
Mimegralla coeruleifrons	678	Karanganyar	This research
Hemichaeta scutellata	1493	CR: Dominical Hacienda Baru	Jackson <i>et al</i> . (2014) KM28700.1
Taeniaptera trivittata	1376	USA: SC: Georgetown Co.	Jackson <i>et al.</i> (2014) KM287302.1
Micropeza sp.	1405	Argentina	Jackson <i>et al.</i> (2014) KM287330.1

Table 3. Origin of *Mimegralla* spp. and three other species of Micropezidae Family from GenBank database were used for phylogenetic comparison

 Table 4. Percentage of homology from nucleotide base of Mimegralla albimana and three others species of Micropezidae Family from GenBank database

No.	Species	1	2	3	4	5
1	Mimegralla albimana	ID				
2	Mimegralla coeruleifrons	88%	ID			
3	Hemichaeta scutellata	88%	85%	ID		
4	Taeniaptera trivittata	87%	84%	88%	ID	
5	Micropeza sp.	87%	84%	86%	89%	ID



Figure 5. Phylogeny tree of Mimegralla spp. and others species of Micropezidae Family

could be determined by counting nucleotide base without an asterisk (*). The result indicated that M. coeruleifrons has closest relationship with M. albimana (88%) than with the other three species (Hemichaeta scutellata, Taeniaptera trivittata, and Micropeza sp.) on homology (Table 4) and clustal consensus (Figure 4). Sokal and Sneath (1963) stated that the closeness of phylogenic relationship between organisms are indicated by the similarity of common characters. Figure 5 showed that M. coeruleifrons and M. albimana has a bootstrap value at 92, means that these species are member of Mimegralla Genus. Bootstrap analysis is used to test the validity and reliability of data (Hedges, 1992). Thus, the value of 92 means that the data in phylogeny analysis has high validity and reliability.

CONCLUSION

M. coeruleifrons and *M. albimana* found in Zingiberaceae plants (ginger, turmeric, javanese ginger, and aromatic ginger) were member of the same genus, Mimegralla, by the facts that these two species have high values of phylogenic relationship (88%) and bootstrap (92).

ACKNOWLEDGEMENTS

The author would like to thank Prof. Steve Marshall, Dr. Awit Suwito and Dr. Rodiah Balfas for identification assistance, Siti Haryati for DNA extraction assistance, Abdu Rohman and Rizki Rajabillah for phylogeny discussion. Funding for this project was provide by BPPSDM of Ministry of Health.

LITERATURE CITED

Aczel, M.L. 1959. *Insects of Micronesia, Diptera: Neriidae and Micropezidae*. Bernice P. Bishop Museum, Hawaii. 90 p.

Balfas, R. 2002. Status Lalat Rimpang pada Tanaman Jahe dan Strategi Pengendaliannya (Status of Rhizome Fly at Ginger and their Control). *Jurnal Litbangtan (Journal of Research and Development on Agriculture)* 21: 32–37.

Balfas, R., Supriadi, N. Karyani, & E. Sugandi. 2000. Serangan *Mimegralla coeruleifrons* Macquart dan Peranannya dalam Membawa Patogen Bakteri Penyakit Layu (Attack of *Mimegralla coeruleifrons* Macquart and its Role in Bringing Pathogen of Wilt Disease. *Jurnal Penelitian Tanaman Industri* (*Research Journal of Industrial Plant*) 5: 123–127.

Folmer, O., M. Black, W. Hoeh, R. Lutz, & R. Vrijenhoe. 1994. DNA Primers for Amplification of Mitocondrial Cytochrom C Oxidase Subunit I from Diverse Metazoan Invertebrates. *Molecular Marine Biology and Biotechnology* 3: 294–299.

Ghorpade, S.A., S.S. Jadhav, & D.S. Ajri. 1983. Survey of Rhizome Fly on Turmeric and Ginger in Maharashtra. *Journal Maharashtra Agriculture University* 8: 292–293.

Ghorpade, S.A., S.S. Jadhav, & D.S. Ajri. 1988. Biology of Rhizome Fly, *Mimegralla coeruleifrons* Macquart (Micropezidae: Diptera) in India, a Pest of Turmeric and Ginger Crops. *Tropical Pest Management* 34: 48–51.

Hedges, S.B. 1992. The Number of Replications Needed for Accurate Estimation of the Bootstrap p-Value in Phylogenetic Studies. *Molecular Biology and Evolution* 9: 366–369. Hennig, W. 1935. Revision der Tyliden (Dipt., Acalypt.) II Teil: Die außeramerikanischen Taeniapterinae, die Trepidariinae und Tylinae. Allgemeines über die Tyliden. (Fortsetzung). *Konowia* 14: 192–216.

Joy, P.P., J. Thomas, S. Mathew, & B.P. Skaria. 1998. *Zingiberaceous Medicinal and Aromatic Plants*. Aromatic and Medicinal Plants Research Station, Odakkali, Asamannoor P.O., Kerala, India. 31p.

Karmawati, E., B. Baringbing, M. Iskandar, & T.E. Wahyono. 1990. Observasi Lalat Rimpang pada Pertanaman Jahe di K.P. Sukamulya. (Observation of Rhizome Fly on Ginger Plants at Sukamulya Village). *Media Komunikasi Penelitian dan Pengembangan Tanaman Industri (Media Communication Research and Development Industrial Crops*) 6: 84–86.

Maxwell-Lefroy, H. & F.M. Howlett. 1909. *Indian Insect Life-A Manual of the Insects of the Plains (Tropical India)*. Government Press, Calcutta. 486 p.

Nair, M.R.G.K. 1980. Pests of Ginger and Turmeric, p. 101–103. *In* M.K. Nair, T. Premkumar, P.N. Ravindran, & Y.R. Sarma (eds.), *Proceedings of the National Seminar on Ginger and Turmeric*. Calicut, 8–9 April 1980.

Pape, T., V. Blagoderov, & M.B. Mostovski. 1758. Order Diptera Linnaeus. *In* Z.Q. Zhang (ed.), Animal Biodiversity: An Outline of Higher-Level Classification and Survey of Taxonomic Richness. *Zootaxa* 3148: 222–229.

Sokal, R.H. & P.A. Sneath. 1963. *Principle of Numerical Taxonomy*. W.H. Freeman and Co., San Fransisco. 359 p.

Steyskal, G.C. 1964. Larvae of Micropezidae (Diptera) Including the Spesies that Bore Ginger Root. *Annal* of the Entomological Society of America 57: 292–296.