Fixed-Bed Column Studies for the Removal of Congo Red Using *Simmondsia chinesis* (Jojoba) and Coated with Chitosan

Amina Abdel Meguid Attia*, Mona Abdel Hamid Shouman, Soheir Abdel Atty Khedr, and Nevin Ahmed Hassan

National Research Center, Laboratory of Surface Chemistry and Catalysis, 33 El-Bohouth St., Dokki, Giza, Egypt, P.O.12622

Received October 23, 2017; Accepted November 20, 2017

ABSTRACT

The goal of this article describes the potential of utilizing jojoba leaves and also modified with chitosan as an efficient adsorption materials for Congo red dye removal in a fixed-bed column. Inlet dye concentration, feed flow rate and bed height had a great influence on determining the breakthrough curves. The percentage dye removal was found to be approximately 69% of coated jojoba leaves with flow rate 3 mL/min, initial concentration 50 mg/L and 4 cm bed height. The dye uptake capacity at equilibrium ($q_e$) for coated jojoba leaves showed higher values than that found for jojoba leaves. On this basis, this implies that the amino groups played an important role during the adsorption process. Breakthrough curves were satisfactorily in good agreement with both Thomas and Yoon-Nelson models based on the values of correlation coefficient ($R^2 \geq 96$). This study serves as a good fundamental aspect of wastewater purification on jojoba leaves as a novel adsorbent for the uptake of Congo red dyes from aqueous solution in a column system.

Keywords: jojoba leaves; adsorption; Congo red; chitosan; fixed-bed column

INTRODUCTION

Lately, industrial developments and its products have the power to enhance and improve the environment community. Dyes and pigments are used in various textile industries to color their products thus leading to the contamination of wastewater [1]. The breakdown of these products influence the life forms causing toxicity to aquatic life [2] and carcinogenic, mutagenic diseases that lead to severe damage to the human beings [3]. Congo red (CR) is a benzidine-based azo dye metabolize to benzidine which generates from plastic industries, rubber printing, and dying etc. The structural stability of Congo red makes it not easily degradable. Treatment of textile wastewater is considered as an environmental challenge and government legislation. As a consequence, there is a grand demand to have an effectual process for the uptake of these dyes [4]. Recently, adsorption using agricultural wastes has been the most economical process for the discharge of various contaminants [5] and heavy metals [6].

*Corresponding author.
Email address: amina_abdelmeguid@yahoo.com
DOI: 10.22146/ijc.29264
reported to have the potential as a novel adsorbent for dye removal [7].

Natural polysaccharide chitosan, a derivative of chitin [8-9] has been known for its unique properties due to the existence of high content of amino and hydroxyl functional groups in its molecules that contribute in the process of adsorption [10]. Moreover, chitosan is a nontoxic, biodegradable, and biocompatible polymer, which can be used as a wound dressing, tissue engineering, and drug delivery systems [11]. However, chitosan has unconventional properties that make it become limited to use for adsorption such as limited specific gravity, simplicity in agglomeration, poor mechanical strength in addition to lack of solubility in aqueous acids [12]. Therefore, chitosan is often used to combine with other materials to enhance their extraordinary properties. Several attempts were carried out to give desirable mechanical and effective adsorption through binding chitosan with other materials and using it for hazardous materials as well as dye uptake. Jojoba leaves are incorporated with chitosan to create an adsorbent that improves the properties of chitosan. In addition, it exhibits a porous structure that makes it become widely used as an adsorbent for removal of pollutants. Recently, pretreatment of the surface by amine groups could affect the properties of the sorbent and enhance its adsorption capacity for anionic dyes [13]. Batch adsorption experiments are used easily in the lab for the remedy of a small volume of effluents but less convenient to utilize on an industrial scale, where large quantities of wastewater are produced. However, batch adsorption affords fundamental knowledge about the efficiency of the sorbent-sorbate system. Furthermore, the data gained by the batch condition is not suitable for the most treatment systems (such as column studies) in which the contact time is not sufficient to reach equilibrium [14]. Thus, it is needed to find out the practical applicability of the sorbent in the continuous mode. In fact, the fixed-bed adsorption study is a feasible solution for the detection of bed operation lifespan, regeneration time and determines the experimental breakthrough curve [15].

The article provides a new approach to study the potential of Jojoba leaves and that coated with chitosan for the uptake of Congo red in a fixed- bed column. Operating variables such as initial dye concentration, bed height, and flow rate were considered. The results were applied to Thomas and Yoon-Nelson models.

**EXPERIMENTAL SECTION**

**Materials**

Raw Jojoba leaves (JL) were collected from trees planted in a number of deserts predominately in Southern Egypt. The leaves were washed thoroughly with distilled water to remove dirt, dust, and other unwanted particles. Afterwards, they were dried in an oven at 60 °C for 24 h. The dried samples were milled and passed through a molecular sieve of 0.6 mm.

**Instrumentation**

Congo red concentrations of solutions before and after adsorption were measured using a UV-Vis Spectrophotometer (UV-2401, Shimadzu) at a wavelength of 498 nm which corresponds to the maximum absorbance of the dye. The Fourier Transform Infrared absorption spectra (FT-IR) was recorded on KBr pressed pellets of the powdered sample in the range 4000-400 cm⁻¹ using a Perkin Elmer FTIR 2000 Spectrophotometer. The surface morphology of the samples was investigated using scanning electron microscopy (SEM, JOEL Model JSM 5300LV). Images were captured in a liquid nitrogen cooled atmosphere.

**Procedure**

**Preparation of dye solution**

The Congo red (CR) obtained from Fluka was used without any purification. The chemical formula of CR is C36H22N4Na2O8S6 with a molecular weight 696.66 g/mol. The chemical structure of the dye is shown in Fig. 1. The stock dye solution (100 mg/L) was prepared by dissolving 0.1 g of CR in 1000 mL distilled water. Experiments were carried out by diluting the stock solutions to the desired concentration.

**Preparation of chitosan gel**

Chitosan (2-Amino-2-deoxy-(1→4)-β-D-glucopyranose) was supplied by Sigma-Aldrich chemical company. About 20 g of chitosan was slowly added to 10% (v/v) glacial acetic acid with constant stirring for 2 h at 40–50 °C to produce a whitish viscous gel.
Coating with Chitosan

Chitosan coating process on the surface of Jojoba leaves (CJL) is done by soaking 25 g of Jojoba in 50 mL chitosan gel and then heated at a temperature of 40–50°C for 1 h with continuous stirring then dried in an oven at 100°C overnight.

Fixed-bed adsorption studies

Continuous flow adsorption experiments were conducted in a 2 cm internal diameter and 50 cm height glass column at room temperature. Three parameters including initial concentration (50 and 100 mg/L), flow rate (3 and 5 mL/min) and bed height (3 and 4 cm) were studied in the column test. The column was filled with adsorbate between two graded levels of quartz sand to avoid the loss of adsorbent and also to ensure a closely packed arrangement. In order to eliminate air bubbles inside the column, a definite volume of distilled water was passed through the column prior to adsorption study. Dye solution of known concentration was pumped downward through the column by a peristaltic pump. Samples were collected at the outlet of the column at regular time intervals and the concentrations of dye in the effluent were analyzed using UV/VIS spectrophotometer.

Fixed-bed column data analysis

Adsorption characteristics in a fixed-bed column were studied by analyzing the shape of the experimental breakthrough curves. Breakthrough curves are expressed in term of C/C0 as a function of time for a given condition, where C is the concentration of influent and C0 is the concentration of effluent [16]. The exhaustion point is determined when C changes to 95% of C0. The effluent volume (Ve) can be calculated as eq. 1:

\[ V_{e} = Q \int_{0}^{t} C_{e} \, dt \]  

where \( V_{e} \) is the effluent volume collected (mL); \( Q \) is the volumetric flow rate (mL/min), and \( t \) is the total flow time (min).

The total amount of dye adsorbed for a given flow rate, feed concentration, and bed height is given by the eq. 2:

\[ q_{\text{total}} = \frac{Q}{1000} \int_{0}^{t} C_{e} \, dt \]  

where \( q_{\text{total}} \) is the maximum bed capacity (mg); \( Q \) is the inlet flow rate (mL/min); \( C_{e} \) is adsorbed dye concentration (mg/L). The value of the integral is obtained from the area under the curve of adsorbed dye concentration versus time. Besides, the equilibrium uptake \( q_{e} \) or maximum capacity of the column can be calculated as eq. 3:

\[ q_{e} = \frac{q_{\text{total}}}{W} \]  

where \( q_{\text{total}} \) is the amount of dye adsorbed (mg) and \( W \) is the weight of adsorbent (mg).

The total amount of dye is given by the following eq. 4:

\[ M_{\text{total}} = \frac{C_{0} Q t_{\text{total}}}{1000} \]  

where \( M_{\text{total}} \) is the total amount of dye (g); \( C_{0} \) is the initial dye concentration (mg/L); \( Q \) is the volumetric flow rate (mL/min); and \( t_{\text{total}} \) is the total flow time (min).

The percentage removal of dye is given as eq. 5:

\[ \% \text{Removal} = \frac{q_{\text{total}} \times 100}{M_{\text{total}}} \]  

where \( q_{\text{total}} \) is the maximum bed capacity (mg); \( M_{\text{total}} \) is the amount of dye sent to the column (mg).

Modeling of breakthrough curves

Various mathematical models are used to describe the fixed-bed column behavior. In many cases, kinetics of adsorption in a column have been analyzed for Thomas and Yoon-Nelson model to give a suitable description of the adsorption kinetics. Thus, in this research, a trial has been made to find the best model defining the adsorption kinetics to study the breakthrough performance and to determine the adsorption capacity of the fixed bed column.

Thomas model [17] is based on the mass transfer model which postulates that dye emigrates from the solution to a film around the particle and expands through the liquid film to the surface of the adsorbent. Subsequently, this step is followed by intraparticle diffusion and adsorption on the active site. Its assume Langmuir isotherm for equilibrium, plug flow performance in the bed and second-order reversible reaction kinetics. The linear form of the Thomas model is expressed by the eq. 6:

\[ \ln \left( \frac{C_{0}}{C_{t}} - 1 \right) = K_{TH} t = XQ - K_{TH} C_{t} t \]  

where \( C_{0} \) is the initial dye concentration (mg/L); \( C_{t} \) is effluent dye concentration at time \( t \) (mg/L); \( K_{TH} \) is Thomas model constant (L/min.mg); \( Q \) is the maximum adsorption capacity (mg/g); \( X \) is the mass of adsorbent (g); \( Q \) is inlet flow concentration (mL/min). The value of \( K_{TH} \) and \( Q \) are determined from the slope and intercept of a plot of \( \ln \left( C_{0}/C_{t} - 1 \right) \) versus \( t \).

The Yoon-Nelson model [18] is based on the assumption that the rate of decrease in the probability of adsorption for each adsorbate molecule is proportional to the probability of adsorbate adsorption and the probability of adsorption breakthrough on the adsorbent. The linear form of Yoon-Nelson model is given by the eq. 7:

\[ \ln \left( \frac{C_{0}}{C_{t} - C_{e}} \right) = K_{YN} t - K_{YN} t \]  

where \( C_{t} \) is the effluent dye concentration, \( C_{0} \) is the initial dye concentration, \( C_{e} \) is adsorbed dye concentration, and \( t \) is time (min).

Amina Abdel Meguid Attia et al.
where $C_0$ is the initial dye concentration (mg/L); $C_t$ is dye concentration at time $t$ (mg/L); $t$ is the flow time (min); $\tau$ is the time required for 50% breakthrough (min); $K_{YN}$ is Yoon-Nelson rate constant (min$^{-1}$). These values $K_{YN}$ and $\tau$ are determined from the slope and intercept of $\ln (C_t/C_0-C_t)$ versus $t$. The model parameters were determined by the fit of models with the experimental data through nonlinear regression. The fit quantity was measured through the coefficient of determination ($R^2$).

RESULT AND DISCUSSION

Characterization of Adsorbent

Scanning electron microscope analysis

The morphological structure of (JL) and (CJL) were presented in Fig. 2. It reveals that the jojoba leaves possess a rough surface, non-homogeneous with significant remarkable irregularities. However, coating with chitosan led to the formation of some wrinkles that is less porous which suggests that the pores could have acted as active sites in the dye adsorption. Furthermore, the surface was accompanied by fading appearance [19].

Fourier transform infrared spectroscopy analysis

FTIR analysis of (JL) and (CJL) was performed in order to describe the functional groups responsible for the binding mechanism between the adsorbent and the coated sample. Both samples demonstrated similar characteristic peaks while minute changes arise during the coating of Jojoba leaves as presented in the Fig. 3. The FTIR spectrum of (JL) shows a predominant broadband around 3238 cm$^{-1}$ which correspond to the stretching vibration of O–H groups. The bands around 2921 and 2852 cm$^{-1}$ were assigned for asymmetric and symmetric stretching vibration of CH groups [20]. A large cluster of functional groups like $\text{–C=O, –COO}$ were found at wave number intervals of 1626 and 1457 cm$^{-1}$. Common adsorption band of $\text{–CH}$ around wave numbers 1626 and 1027 cm$^{-1}$ was shifted after coating the leaves. The peak at 1737 and 1527 cm$^{-1}$ was attributed to the presence of oxygen functional groups such as a conjugated C–O stretching in carboxylic groups [21]. Another absorption bands revealing around 1238 and 452 cm$^{-1}$ contributed to the Si–C stretch and C–O–H twist [22]. The observed peak at 1027 cm$^{-1}$ is assigned to $\text{–CN}$ stretch of aliphatic amines [23]. On interpreting the FTIR spectra of chitosan-coated Jojoba leaves, the spectrum displays the absorption of both chitosan and the raw material. The major bands arising in the spectrum were as a result of the stretching vibrations of $\text{–OH}$ groups in the range 3750 to 3000 cm$^{-1}$ which are imbricate to the stretching vibration of N–H and C–H band in CH$_2$ (2922 cm$^{-1}$) and $\text{–CH}_2$ (2852 cm$^{-1}$) groups, respectively [24]. Absorption band in the range of 1627–1450 cm$^{-1}$ was ascribed to the vibration of carbonyl bonds (C=O) of the amide group CONHR [secondary amide] and the vibration of the protonated amine group. The limited peak at 515 cm$^{-1}$ correlates to oscillating of the saccharide structure of chitosan [25].

X-ray fluorescence analysis

The mineralogical composition of pure chitosan and investigated adsorbents were depicted in Table 1. The major constituents detected by the XRF of pure chitosan and the Jojoba leaves are mainly Si, Fe, Al, and Ca. After the coating process, the percentage of these elements increased. Therefore, these findings were quite surprising and suggest the possibility that chitosan could be successfully cross-linked on the surface of Jojoba leaves.

![Fig 2. SEM image of (a) JL and (b) CJL](image-url)
Fig 3. FTIR spectra of (a) JL and (b) CJL

Table 1. Chemical composition of chitosan, Jojoba leaves and coated jojoba leaves

<table>
<thead>
<tr>
<th>Main Constituents (wt.%)</th>
<th>Chitosan</th>
<th>JL</th>
<th>CJL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Si</td>
<td>1.373</td>
<td>0.079</td>
<td>0.134</td>
</tr>
<tr>
<td>Al</td>
<td>0.106</td>
<td>0.024</td>
<td>0.036</td>
</tr>
<tr>
<td>Fe</td>
<td>0.091</td>
<td>0.061</td>
<td>0.092</td>
</tr>
<tr>
<td>Mn</td>
<td>0.016</td>
<td>ND</td>
<td>0.008</td>
</tr>
<tr>
<td>Zn</td>
<td>ND</td>
<td>0.007</td>
<td>0.005</td>
</tr>
<tr>
<td>Sr</td>
<td>ND</td>
<td>0.002</td>
<td>0.003</td>
</tr>
<tr>
<td>Mg</td>
<td>ND</td>
<td>0.127</td>
<td>0.096</td>
</tr>
<tr>
<td>Ca</td>
<td>66.260</td>
<td>0.233</td>
<td>0.282</td>
</tr>
<tr>
<td>K</td>
<td>ND</td>
<td>0.495</td>
<td>0.480</td>
</tr>
<tr>
<td>Na</td>
<td>ND</td>
<td>0.049</td>
<td>0.044</td>
</tr>
<tr>
<td>Nd</td>
<td>ND</td>
<td>ND</td>
<td>0.024</td>
</tr>
<tr>
<td>P</td>
<td>ND</td>
<td>0.249</td>
<td>0.277</td>
</tr>
<tr>
<td>S</td>
<td>ND</td>
<td>0.346</td>
<td>0.353</td>
</tr>
<tr>
<td>Cl</td>
<td>ND</td>
<td>0.136</td>
<td>0.173</td>
</tr>
<tr>
<td>LO</td>
<td>67.846</td>
<td>98.190</td>
<td>97.990</td>
</tr>
</tbody>
</table>

ND = not detected
Fig 4. Effect of initial dye concentration on breakthrough curve for fixed-bed adsorption of CR by (a) JL and (b) CJL

Table 2. Column parameters of raw Jojoba leaves for CR adsorption at different initial concentration, bed height, and flow rate

<table>
<thead>
<tr>
<th>Initial concentration</th>
<th>t_{total} (min)</th>
<th>q_{total} (mg)</th>
<th>q_{exp} (mg/g)</th>
<th>M_{total} (g)</th>
<th>% Removal</th>
</tr>
</thead>
<tbody>
<tr>
<td>50 mg/L</td>
<td>200</td>
<td>13</td>
<td>0.004</td>
<td>30</td>
<td>57</td>
</tr>
<tr>
<td>100 mg/L</td>
<td>150</td>
<td>26</td>
<td>0.008</td>
<td>45</td>
<td>43</td>
</tr>
<tr>
<td>Bed height</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 cm</td>
<td>150</td>
<td>20</td>
<td>0.003</td>
<td>45</td>
<td>46</td>
</tr>
<tr>
<td>4 cm</td>
<td>211</td>
<td>40</td>
<td>0.009</td>
<td>63</td>
<td>57</td>
</tr>
<tr>
<td>Flow rate</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 mL/min</td>
<td>150</td>
<td>21</td>
<td>0.003</td>
<td>45</td>
<td>57</td>
</tr>
<tr>
<td>5 mL/min</td>
<td>120</td>
<td>14</td>
<td>0.004</td>
<td>50</td>
<td>29</td>
</tr>
</tbody>
</table>

Table 3. Column parameters of coated Jojoba leaves for CR adsorption at different initial concentration, bed height, and flow rate

<table>
<thead>
<tr>
<th>Initial concentration</th>
<th>t_{total} (min)</th>
<th>q_{total} (mg)</th>
<th>q_{exp} (mg/g)</th>
<th>M_{total} (g)</th>
<th>% Removal</th>
</tr>
</thead>
<tbody>
<tr>
<td>50 mg/L</td>
<td>222</td>
<td>18</td>
<td>0.006</td>
<td>33</td>
<td>69</td>
</tr>
<tr>
<td>100 mg/L</td>
<td>185</td>
<td>35</td>
<td>0.014</td>
<td>56</td>
<td>56</td>
</tr>
<tr>
<td>Bed height</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 cm</td>
<td>185</td>
<td>35</td>
<td>0.014</td>
<td>56</td>
<td>56</td>
</tr>
<tr>
<td>4 cm</td>
<td>350</td>
<td>73</td>
<td>0.020</td>
<td>105</td>
<td>69</td>
</tr>
<tr>
<td>Flow rate</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 mL/min</td>
<td>185</td>
<td>35</td>
<td>0.014</td>
<td>56</td>
<td>69</td>
</tr>
<tr>
<td>5 mL/min</td>
<td>150</td>
<td>26</td>
<td>0.008</td>
<td>75</td>
<td>35</td>
</tr>
</tbody>
</table>

Fixed-Bed Adsorption Studies

Effect of initial dye concentration

The performance of the breakthrough curves is illustrated in Fig. 4 by using different dye concentrations. From this figure, it was obvious that the breakthrough occurred in a short period of time for high dye concentration, while the breakthrough occurred latterly for low dye inlet concentration and the surface of the adsorbents was saturated after a long period of time. Therefore, at a lower concentration, the breakthrough was planer due to lower mass transfer in the adsorption process, on the contrary, at high inlet dye concentration the profile of breakthrough were sharp implying an intra-particle diffusion process. These results can be interpreted on the basis that further adsorption sites were shielded by increasing the dye concentration and the driving force for the transfer process will overcome the mass transfer resistance. Therefore, at low inlet concentration, the binding sites of the adsorbents interact with every dye molecule exists in the solution thus permitting higher adsorption process. While, for higher concentration, excess molecules of the dye were unadsorbed which led to a decrease in the adsorption process [26]. The column parameters obtained from the effect of initial dye concentrations for JL and CJL are given in Table 2 and 3. As can be seen, the total amount of CR dye adsorbed increased,
while the percentage removal decreased for both JL and CJL. However, the amount of CJL showed the highest removal.

**Effect of flow rate**

The flow rate is an essential factor for predicting the proficiency of adsorbent. The breakthrough curves at various flow rates are presented in Fig. 5. This figure displays that breakthrough curves mostly occurred faster at a higher flow rate which may be correlated with the availability of reaction sites able to capture dye molecules inside or around the adsorbents, so the removal efficiency became ineffective. Additionally, the column was capable to accumulate CR dye after the occurring of breakthrough curve. It has been depicted that increasing the flow rate was accompanied by a decrease in dye removal. This interpretation was based on the fact that the residence time of CR dye was not sufficient to reach adsorption equilibrium at that flow rate. This can be explained on the basis that there is not enough contact time between the dye and the adsorbent resulting in less diffusion of liquid present in the column.

This induces a lower distribution of the solute through the adsorbent [27]. The column parameters obtained from the effect of flow rate are listed in Table 2 and 3. As can be seen, the total amount of CR dye adsorbed increased, while the percentage removal decreased.

**Effect of bed depth**

The quantity of adsorbent inside the column greatly affects the accumulation of adsorbate in the fixed-bed column. Fig. 6 presents the effect of bed height on simulated breakthrough curves. It was revealed from this figure, that by increasing the bed height, this allows the solute to have enough time to diffuse into both adsorbents and permits high removal efficiency. The slope of the breakthrough curve decreased with an increase in the bed height which derived in a broadened mass transfer zone. This was owed to a large surface area of adsorbents leading to more binding sites for adsorption [28-29]. The column parameters for both adsorbents obtained from the effect of bed height are given in Table 2 and 3.
All Tables display that the removal capacity of CR by coated jojoba leaves showed higher values over that of raw Jojoba. Based on these results, it can be ascribed that the adsorption process is an electrostatic process between the anionic dye (CR) and the amino groups of the chitosan. In aqueous solutions, CR dye is ionized into anionic dye with sulphonate groups and the amino groups were protonated which makes it positively charge. Thus, the removal of CR with chitosan is mainly due to the electrostatic reaction between the amino groups of chitosan (NH\(^3\)) and the sulphonate group of the anionic dye (SO\(_3^-\)).

**Breakthrough Curve Modeling**

Several mathematical models have been advanced for analyzing column performance and to scale it up for industrial utilization. In this research, Thomas and Yoon-Nelson models were applied to identify the best model for predicting the dynamic behavior of the column.

**Thomas model**

The column data were fitted to the Thomas model to determine the constants \(K_{TH}\) and \(q_0\) which were calculated from the slope and intercept of a linear plot of \(\ln (C_o/C_t - 1)\) versus time under different adsorption parameters (flow rate, dye concentration, and bed height) for both adsorbents. The prediction of breakthrough curves for JL and CJL conferring to Thomas model is represented in Fig. 7 and 8. The obtained Thomas model parameters and experimental values of \(q_o\) are provided in Table 4. According to the determined coefficient (\(R^2\)), Thomas model was applicable to the experimental data. In addition, the consistent values of \(q_o\) and \(q_{oexp}\) displayed the suitability of this model. Increasing the influent concentration led to an increase in the value of \(K_{TH}\) and a decrease in the value of \(q_o\). This is attributed to the driving force of adsorption between the dye existed on both adsorbents and the dye present in the solution [30-31]. However, \(q_0\) increased and \(K_{TH}\) decreased with...
Fig 8. Thomas kinetic plot for the adsorption of CR on CJL at different (a) flow rate (b) bed depth and (c) initial concentration

Table 4. Column kinetic parameters for the adsorption of CR using Thomas and Yoon - Nelson Models

<table>
<thead>
<tr>
<th>Adsorbents</th>
<th>Flow rate (mL)</th>
<th>K_TH(mL/min/mg)</th>
<th>q_o (mg/g)</th>
<th>R²</th>
<th>K_YN(min⁻¹)</th>
<th>T</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>JL</td>
<td>3</td>
<td>0.030</td>
<td>5.90</td>
<td>0.99</td>
<td>0.042</td>
<td>130</td>
<td>0.97</td>
</tr>
<tr>
<td>JL</td>
<td>5</td>
<td>0.040</td>
<td>3.10</td>
<td>0.98</td>
<td>0.056</td>
<td>75</td>
<td>0.98</td>
</tr>
<tr>
<td>CJL</td>
<td>3</td>
<td>0.014</td>
<td>3.08</td>
<td>0.97</td>
<td>0.016</td>
<td>255</td>
<td>0.99</td>
</tr>
<tr>
<td>CJL</td>
<td>5</td>
<td>0.025</td>
<td>1.78</td>
<td>0.98</td>
<td>0.025</td>
<td>105</td>
<td>0.97</td>
</tr>
<tr>
<td>Adsorbents</td>
<td>Initial conc. (mg/L)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>JL</td>
<td>50</td>
<td>0.047</td>
<td>4.60</td>
<td>0.97</td>
<td>0.020</td>
<td>150</td>
<td>0.96</td>
</tr>
<tr>
<td>JL</td>
<td>100</td>
<td>0.030</td>
<td>5.90</td>
<td>0.99</td>
<td>0.042</td>
<td>130</td>
<td>0.97</td>
</tr>
<tr>
<td>CJL</td>
<td>50</td>
<td>0.020</td>
<td>2.50</td>
<td>0.95</td>
<td>0.011</td>
<td>290</td>
<td>0.96</td>
</tr>
<tr>
<td>CJL</td>
<td>100</td>
<td>0.014</td>
<td>3.08</td>
<td>0.97</td>
<td>0.016</td>
<td>255</td>
<td>0.99</td>
</tr>
<tr>
<td>Adsorbents</td>
<td>Bed Height (g)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>JL</td>
<td>3</td>
<td>0.030</td>
<td>5.90</td>
<td>0.99</td>
<td>0.042</td>
<td>130</td>
<td>0.97</td>
</tr>
<tr>
<td>JL</td>
<td>4</td>
<td>0.025</td>
<td>6.50</td>
<td>0.98</td>
<td>0.037</td>
<td>150</td>
<td>0.99</td>
</tr>
<tr>
<td>CJL</td>
<td>3</td>
<td>0.014</td>
<td>3.08</td>
<td>0.97</td>
<td>0.016</td>
<td>255</td>
<td>0.99</td>
</tr>
<tr>
<td>CJL</td>
<td>4</td>
<td>0.009</td>
<td>4.40</td>
<td>0.98</td>
<td>0.009</td>
<td>360</td>
<td>0.98</td>
</tr>
</tbody>
</table>

the increase in bed height. This might be attributed to the increase in mass-transport resistance with increasing bed height in columns. Furthermore, with increasing the flow rate, the value of q_o decreased but the value of K_TH increased. This is due to that at higher flow rate there is a deficient residence time of the solution inside the column and migration on of the solute into the pores of both adsorbents which leads to lower adsorption capacity and thus the solute leaves the column before equilibrium occurs [32]. Accordingly, with lower flow rate, lower initial dye concentration and higher bed depth would increase the adsorption of CR on both adsorbents column. The experimental data were fitted with the linear form of the Thomas model indicates the absence of axial dispersion where the exterior and interior diffusion is not the limiting step.
**Fig 9.** Yoon - Nelson kinetic plot for the adsorption of CR on JL at different (a) flow rate (b) bed depth and (c) initial concentration

**Fig 10.** Yoon - Nelson kinetic plot for the adsorption of CR on CJL at different (a) flow rate (b) bed depth and (c) initial concentration
Yoon-Nelson model

Yoon-Nelson Model was applied to predict the adsorption technique and the breakthrough curves. The values of $\tau$ and $K_{YN}$ were calculated from the linear plots of $\ln (C_i/C_e - C)$ versus time (min) at different flow rates, bed heights and initial dye concentrations for the adsorption of CR onto JL and CJL (Fig. 9 and 10) and their values are listed in Table 4. It is obviously shown that as the initial dye concentration and flow rate increased for both adsorbents, the value of $\tau$ decreased and $K_{YN}$ increased. This is owing to the minor residence time of solute in the column. Additionally, the force controlling the mass transfer in liquid phase could clarify the increase in $K_{YN}$. This led to a prolonged time for the bed to get saturated. However, increasing the bed height led to a decrease in the in the rate constant $K_{YN}$ and $\tau$ increased. This result is ascribed to the availability of more adsorption sites [33-34]. It can be seen according to the values of correlation coefficient ($R^2$) that Yoon-Nelson model adequately describes the adsorption of CR on both adsorbents to the experimental data. Accordingly, to these findings the coated Jojoba leaves displayed more appropriate adsorption performance than raw Jojoba leaves.

CONCLUSION

This study validated that the raw Jojoba leaves and the modified chitosan were effective adsorbents for the uptake of CR in fixed-bed column studies. FTIR analysis illustrates that chitosan was successfully loaded onto jojoba leaves. Results revealed that the adsorption capacity of the modified leaves was higher than that for jojoba leaves. The dynamic dye adsorption data was studied under varying conditions including initial dye concentration, flow rate, and bed height. From the outcome of our investigation, it is possible to conclude that the percentage dye removal was found to increase by increasing the bed height, while decreased with the increase in flow rate and initial dye concentration. However, increasing bed height resulted in an increase in the breakthrough time while decreased by increasing the dye concentration and flow rate. The revelation of breakthrough curves and the determination of the kinetic column parameters were studied by using the Thomas and Yoon-Nelson models. The high values of correlation coefficients ($R^2$) indicated the applicability of both models.

REFERENCES


