

Journal of the Medical Sciences (Berkala Ilmu Kedokteran)

Volume 52, Number 2, 2020; 181-190 http://dx.doi.org/10.19106/JMedSci005202202010

A fatal acute appendicitis with sepsis and pneumonia was caused by melioidosis: a case report

Abu Tholib Aman^{1,2*}, Yuli Mawarti^{1,2}, Agus Barmawi³, Faisal Heryono⁴, Rizka Humardewayanti Asdie^{2,4}

¹Department of Microbiology, ²Indonesia-Research Partnership on Infectious Disease (INA-RESPOND) Site 580, ³Department of Surgery, ⁴Department of Internal Medicine, Faculty of Medicine, Public Health and Nursing, Universitas Gadjah Mada/Dr. Sardjito General Hospital, Yogyakarta, Indonesia

ABSTRACT

Submited: 2019-12-27 We report anunderdiagnosed fatal case of melioidosis that involved dygestion Accepted : 2020-04-24 system which complicated with pneumonia, and sepsis. The case was initially diagnosed as acute appendicitis, and subsequently the patient underwent an exploratory laparatomy and appendectomy. He was discharged afer 3 days of hospitalization. Thirty days afterward, he was admitted to another private hospital to experience another exploratory laparatomy with indication of pancreatitis, intra-abdominal organs adhesions, and postoperative enterocutaneous fistula (ECF), and hospitalized there for 25 days. He eventually suffered from sepsis, pneumonia, unclosed ECF, anemia, hypoalbuminemia, and electrolyte imbalance. He then referred to a tertiary teaching hospital and hospitalized there for a total 134 days until he passed away. His clinical condition was declining, despite a long course of broad spectrum antibiotics. Treatment delay, prolong hospitalization, and complications were the inevitable, although Burkholderia pseudomallei was finally identified 2 weeks prior to his death. This case highlight that melioidosis canassociate with acute appendicitis, and that the delay on its diagnosis and treatment may trigger complications and death.

ABSTRAK

Kami melaporkan kasus fatal melioidosis yang melibatkan sistem pencernaan dengan komplikasi pneumonia, sepsis, dan berakibat pada kematian. Appendicitis akut adalah diagnosis klinis awal pada kasus melioidosis ini, dan pasien langsung menjalani operasi laparatomi eksplorasi dan appendectomy, kemudian pulang setelah mondok selama 3 hari di sebuah rumah sakit swasta. Tiga puluh hari setelahnya, pasien mondok di rumah sakit swasta lainnya selama 25 hari, dan menjalani operasi laparatomi eksplorasi yang ke dua dengan indikasi pankreatitis, perlengketan organ intra abdomen, dan fistula enterokutan. Kondisi klinis pasien memburuk, dan terjadi sepsis disertai penumonia, luka fistula entero-kutan (FEK) terbuka, anemia, hypoalbuminenia, dan ketidakseimbangan elektrolit. Kemudian pasien dirujuk ke rumah sakit pusat rujukan dan pendidikan dan mondok selama 134 hari sebelum akhirnya meninggal. Kondisi klinis pasien terus memburuk meskipun telah mendapat rangkaian terapi antibiotik berspektrum luas. Keterlambatan terapi, lamanya waktu pemondokan, dan terjadinya komplikasi menjadi tidak terelakkan, meskipun B. pseudomallei dapat diidentifikasi pada 2 minggu sebelum kematian. Kasus ini menekankan pentingnya memahami bahwa presentasi melioidosis secara klinis dapat berhubungan dengan appendicitis akut, dan keterlambatan dalam mendiagnosis dan terapi dapat memicu terjadinya komplikasi dan kematian.

Keywords:

melioidosis; fatal; delay in diagnosing; *B. pseudomallei;* appendicitis;

INTRODUCTION

Melioidosis is an infectious disease caused by Burkholderia pseudomallei.1 Clinical feature of melioidosis vary, with major presentation as sepsis, but specific clinical feature and severity depending on the bacterial entry route into host, host immune response, bacterial strain and load.¹ Case fatality rate of melioidosis range from 10% to 50%, with risk of having recurrent infection is 5-28% in patients who survived acute melioidosis.¹Burkholderia pseudomallei infamous with its diverse antimicrobial resistance feature, including resistant to thirdgeneration cephalosporins, penicillins, rifamycins, and aminoglycosides.² They also showed relative resistance to guinolones and macrolides which limits therapeutic options.² However, most of B. pseudomallei show consistent susceptibility to meropenem and ceftazidim that experts and US CDC considered intravenous meropenem and ceftazidim as effective therapy for melioidosis.^{3,4}

Indonesia, melioidosis In was scarcely reported although some studies predicted that Indonesia was one of endemic countries in Southeast Asia.^{5,6} We described a fatal melioidosis that occurred in 2015. This case report elucidatedthat melioidosis was rarely accurately and timely diagnosed as its clinical presentation might similar or associated with other infectious diaseases, particularly in limited setting like Indonesia. In this case, melioidosis was associated with acute appendicitis and pancreatitis. In addition, multiple attempts of major operation without a timely identification of the etiology, and long term of empirical broad spectrum antibiotics treatment showed a very limited benefit for the patient. This case report highlight the importance of early diagnoses establishment, and appropriate antibiotic management in melioidosis.

CASE REPORT

A 31 years old male suffered from a chronic post surgical wound, after an appendectomy in a private hospital in May 2015. Two weeks after discharged, he complained of abdominal pain, and fever as infection inflicted on the unhealed surgical wound. He went another private hospital, where to multi-slice abdominal computed tomography (MSCT) scan was conducted and revealed a massive gut adhesion and excessive pus surrounding his pancreas. He then was admitted for exploratory laparotomy. However, 25 days after his second operation, an apparent sepsis finally directed his physician to refer him to Dr. Sardjito General Hospital, a tertiary and teaching hospital in Yogyakarta. He was admitted into Emergency

In the emergency department, he appeared skinny (body weight 55 kg, body mass index: 19.03) and weak, with fever (38.6 °C), tenderness, shortness of breath (respiratory rate: 26 cycles/ min), elevated heart rate (148 beat/min), and blood pressure of 107/60 mmHg. The patient chief complain waspain. Pus was continuously dripped from his wound. Laboratory examination supported diagnoses of infection with leucocytosis and neutrophilia (TABLE 1). On physical examination, the attending physicians found consolidation in both side of his lung. A thoracic rontgen photograph showed a prominent right -bilateral pleural effusion (FIGURE 2A). The physicians established diagnosis and management of imminent septic complicated with bilateral shock pneumonia and pleural effusion. anemia, hypoalbuminemia, and surgical site infection of entero-cutaneous fistula (ECF). Resuscitation was conducted in the **Emergency Department**, and antibiotics (ceftriaxone and metronidazole) were administered in a combination.

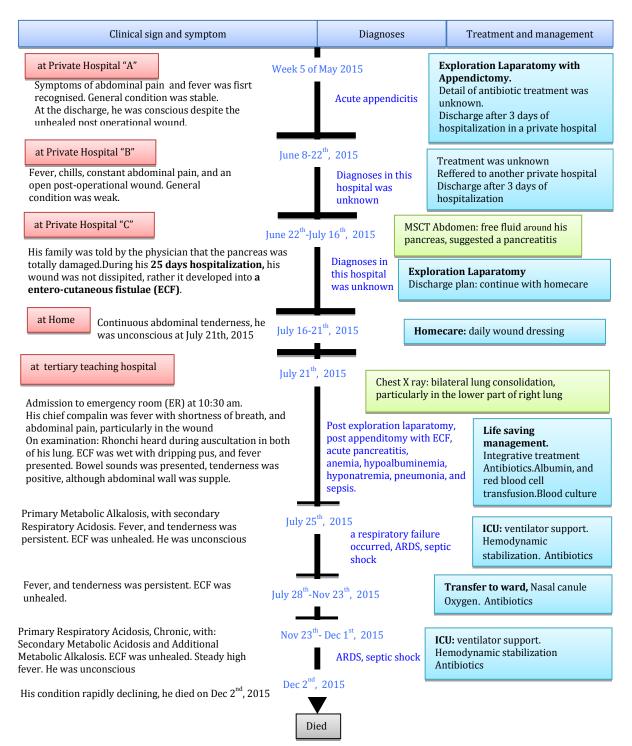


FIGURE 1. A timeline of disease progress, intervention, and outcome of melioidosis.

Test	July 21 th , 2015	July 26 th , 2015 (ICU)	Nov 28 th , 2015 (ICU)	Dec1 st , 2015 (ICU)
Hemoglobine (g/dL)	7.2	7.6	10.4	8.8
Hematocrite (%)	22.9	24.3	32.3	27.9
Leucocyte (/µL)	27,000	19,670	35,160	47,800
Neutrophil (%)	84.4	87	88.7	90.7
Lymphocyte (%)	7.1	6	5.1	4.8
Monocyte (%)	8.4	6.7	3.9	3.7
Basophil (%)	0.1	0.1	0.2	0
Eosinophil (%)	0	0.2	2.1	0.8
Thrombocyte (/µL)	415,000	447,000	281,000	158,000
PH		7.442	7.299	7.295
PO ₂ (mmHg)		105.7	112.3	101,1
PCO ₂ (mmHg)		46.6	51.7	53
SO ₂ (%)		99.6	97.6	96.5
cHCO₃(mmol/L)		29.2	24.8	25.2
BE (mmol/L)		6	-1.9	-1.6
BEecf (mmol/L)		7	-1.6	-1.3
AaDO ₂		484.8	0	336.6
a/AO ₂		17.9	100	23.1
FiO ₂ (%)		0.9	0.21	0.7
Temperature (°C)	38.7	38.3	36.9	37
Albumin (g/L)	1.68	1.65		1.84
Natrium (mmol/L)	131	133	154	153
Chloride (mmol/L)	99	94	114	114
Kalium (mmol/L)	4.3	3.42	3.8	2.9
Amilase	121			
Lipase	180			

TABLE 1. Laboratory tests of the melioidosis case during the hospitalization.

After five days of hospitalization, his condition was worsening. Fever and lung consolidation were persisted although broad-spectrum antibiotics have been administered, and transfusion of albumin and blood, as well as resuscitation had been conducted. On July 25th, 2015 he was transferred into inensive care unit (ICU), as he fell to septic shock. During the 3 days of intensive care, a regimen of antibiotics comprised metronidazole, meropenem, and aztreonam, as well as ventilator support were administed. The patient was transferred back to the ward as his condition was stable on July 27th, 2015. However the following4 months of hospitalization were a prolong fluctuation of clinical condition, which was repeat signs of a temporary stable,pre-shock conditions. The ECF was never healed, and the clinical manifestation was exaggerated with pneumonia, electrolyte imbalance, alkalosis, acidocis, and persistent sign of sepsis, tachypnea, tachycardia, leucocytosis, neutrophilia, lymphopenia, and anemia (TABLE 1).

An abdominal MSCT was conducted at August 7th, 2015 suggested a damaged pancreas, with numerous of small size intra-abdominal granuomas (FIGURE 2B). Amylase and lipase were elevated supported the diagnosis of pancreatitis (TABLE 1), combined with leucocytosis and neutrophilia the suspected etiology was bacterial infection. During the 135 days of hospitalization,nine sets of two sides blood cultures were taken at different days, and the tests were performed with Vitec2, however none of it yielded positive result (TABLE 2). Pus, urine and sputum cultures were also conducted, however it failed to reveal the etiology, until the last wound culture was conducted at day 118 of his hospitalization at another laboratory of microbiology, which revealed *B. pseudomallei* among others bacteria cultivated from the same wound culture (TABLE 2).

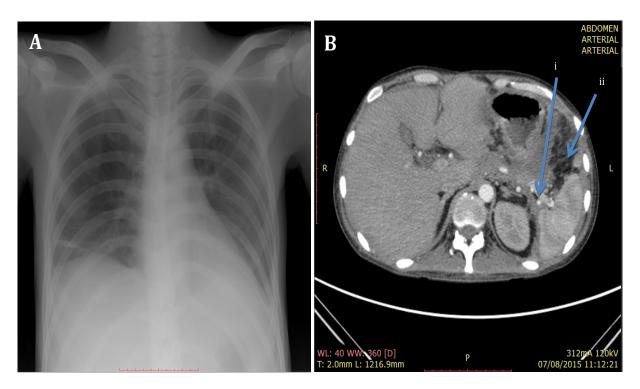


FIGURE 2. Chest X-Ray and abdominal MSCT of the melioidosis. (A). Bilateral lung consolidation was observed from the chest X-ray. (B). MSCT of the abdomen at level of splenic artey (i) and pancreas tail. MSCT showed signs of intraabdominal adhesion, small granulomas (ii).

Antibiotics regimen, steroid, antipyretic were continuously and empirically, following changing response the patient's during hospitalization. In August 2015, he receive antibiotics combination of gentamycin, metronidazole, ceftazidime, levofloxacin, and amikacin, as swell as antifungal fluconazole. The next month, September, the course of antibiotics were amikacin, imipenem, cefixime, and imipenem-cilastatin, with antifungal of fluconazole. In October 2015, imipenem-

cilastatin, levofloxacin, amikacin, and colistin were implemented replaced the previous regimen. In November 2015, the antibiotics regimen comprised imipenem-cilastatin, levofloxacin. cefotaxime, amikacin, ceftazidime, aztreonam, and meropenem. However the clinical condition plunged into septic shock, and at November 24th, 2015 the patient was again transferred to ICU, where he receive intensive care for 8 days before finally he passed away at December 2nd, 2015.

Date	Material	Result
Jul 23 rd , 2015	Blood (2 sites)	Negative
Jul 27 th , 2015	Blood (2 sites)	Negative
Aug 3 rd , 2015	Pus	Negative
Aug 12 th ,2015	Pus	Escherichia coli, ESBL + (contamination?)
Aug 12 th , 2015	Blood (2 sites)	Negative
Aug 16 th , 2015	Blood (2 sites)	Negative
Aug 24 th , 2015	Blood (2 sites)	Negative
Aug 25 th , 2015	Urine	Negative
Aug 29 th , 2015	Sputum	Streptococci
Oct 12 th , 2015	Blood (2 sites)	Negative
Oct 27 th , 2015	Blood (2 sites)	Negative
Nov 3 rd , 2015	Pus	Negative
Nov 11 th , 2015	Blood (1 sites)	Negative
Nov 15 th , 2015	Blood (2 sites)	Negative
Nov 20 th , 2015	Wound swab	 Enterobacter cloacae Klebsiella oxytoca Pseudomonas putida B. pseudomallei , with susceptibility pattern: susceptible to: amikacin, meropenem resistant to: amoxycillin, clavulanate, ampicillin, eritromycin, gentamycin, levofloxacine, chloramphenicol, penicillin, cefepime, cefixime, ceftriaxone, cefuroxime, ciprofloxacine, and sulfamethoxazole

TABLE 2. Culture of blood, pus, urine, sputum, and wound swabs during the
135 days of hospitalization at tertiary teaching hospital.

The research conducted on melioidosis was approved by the Medical and Health Research Ethics Committee, Faculty of Medicine-Public Health and Nursing, Universitas Gadjah Mada, Yogyakarta (Ref.No. KE/FK/1346/ EC/2019). Informed consent was obtained from the next of kin.

DISCUSSION

Melioidosis is a severe infectious disease with diverse clinical manifestations and can be difficult to diagnose as it requires microbiologic expertise to examine the suspected case.⁷ *Burkholderiapseudomallei*isthe causative agent, a gram negative bacteria which able to survive in both intracellular and extracellular for decades in hosts, and endemic to tropical regions of Asia, South America, Central America, Pacific countries, and Indian ocean islands and some African countries.⁷ Burkholderia *pseudomallei*are dubbed as *tropical time* bomb or Vietnamese bomb, as they can cause a latent infection with the longest documented interval between exposure and clinical manifestation is 62 years.⁸ They are tier 1 select agent by the US government, and the culture required special precaution to reduce exposure risk.^{7,8} There is no pathognomonic clinical feature of melioidosis.7 The current standard diagnostic is culture, however the colony of *B. pseudomallei*

be miss-identified can as culture contaminant or other species, especially by laboratory staff who unfamiliar with organism.7 this Burkholderia pseudomallei colonies are usually non-lactose fermenter with metallic sheen and become pink and become umbonated or rugose on Mac Conkey agar after 48 hours.⁷ Under microscope with gram staining, the bacteria appear as bipolar or "safety pin" shaped.⁷ However if the specimen obtained from patients received antibiotics before, the appearance rarely seen, instead *B. pseudomallei* may appear highly atypical resembling yeasts or filamentous.⁷ The one or two days age of colonies usually erroneously dismissed as contaminant or miss-identified as Pseudomonas spp or other organisms when standard diagnosticmethods are used.⁷ The performance or commercially systems widely used such as API 20NE (bioMérieux, Craponne, France), Phoenix (Becton, Dickinson. and Company, Franklin Lakes, NJ, USA), and Vitex 2 (bioMérieux, Craponne, France) to correctly identify B. pseudomallei 37-99%, 0-28%, 0-98% were ranging respectively.7 Moreover, currently no specific and sensitive serologic or rapid tests available to identify *B. pseudomallei*, although experienced laboratory staff can identify them with some additional antibiotic tests and specific media (Ashdown), or confirm them with PCR.⁷Burkholderia pseudomallei is typically susceptible to amoxicillin/ clavulanic acid, trimethoprimsulfamethoxazole, and ceftazidime, but resistant to aminoglycosides, colistin and polymyxin.4,7

Clinical features of melioidosis vary, comprises acute, chronic, and latent diseases. Acute clinical manifestations of melioidosis vary widely, including sepsis, pneumonia, or internal organ abscesses with or without localized infection, therefore melioidosis should be suspected from every patients in endemic regions with community acquired sepsis or pneumonia, urinary tract infection, upper respiratory tract infection, or abscesses especially for those with predisposing factors such as diabetes mellitus, renal disease or immunosuppression.⁷ The chronic manifestations with symptoms usually occur after 2 months of exposure, and are often mimic other diseases such as tuberculosis and cancer.7 The duration of latent melioidosis takes decades, thus a complete travel history should be obtained accordingly.7 Relapse or re-infection with the same or different genotype of *B*. *pseudomallei* may antibiotics treatment.8 occur after Combination of the aforementioned, the lack of clinicians and laboratory staff awareness, and failure to elicit travel history from patients returning from endemic area are contributed to the difficulties on clinical diagnosis of melioidosis.7

The major clinical feature of melioidosis in this case was sepsis, resulted from a prolong misdiagnoses. The reccurent septic shocks were originated from the untreated infection. The unhealed ECF was also a sign of uncontrolled infection. The existed comorbidities and or complication, pneumonia, anemia, such as hypoalbominemia, hiponatremia, and electrolyte imbalance may contribute significantly to the worsening condition and lead to the mortality after long period of hospitalization.

Bacterial culture is the gold standard for diagnose melioidosis.7 However, given the persistent sign and symptoms of infection (high fever although anti-pyretic already administered, leucocytosis, neutrophilia, and monocytosis), blood cultures were unable to identify the etiology. In hyperendemic areas such as Thailand, the positive rate of *B. pseudomallei* from blood culture range from 9.1 to16.5% of suspected melioidosis cases.⁹⁻¹¹ A sevenyear study in Thailand analyzed 63,066 blood cultures reported isolation of *B. pseudomallei* in 11% of 7,296 positive blood cultures.¹²

Melioidosis cases may manifest as systemic infection with cause patient fatality, but blood cultures could result false negative.⁷ Therefore specificmonoclonal antibody assays were suggested to identify melioidosis, as several study reported high sensitivity and specificity (> 95%).^{13,14} In addition, diagnostic tools were rapid also recommended for screening in the area with limited resources where culture is unavailable.¹⁵ Molecular methods are certainly valuable diagnostics tools, which can provide high sensitivity and specificity, as well as rapid identification however the technology is scarcely available in developing countries.¹⁶⁻¹⁸

The major challenge in establishing diagnosis of melioidosis is the selection of colony from culture prior to conducting further identification whether using biochemical or molecular methods.^{19,20} Many bacterial colonies are usually selected after 18-24 h of incubation, since they have produced specific colony features of optimum size of colony.^{7,20} In contrast, the colony of *B. pseudomallei* after 18-24 hours of incubation, appears as a contaminant colony.^{7,20} Experienced laboratory technician will wait up to 3 days to re-evaluate the colony, so that the colony can be identified.^{7,20}

In this case, the problem comprised diagnostic challenges, complications, and the rapid deterioration of patient condition, which unresponsive to the antibiotics treatment. In Yogyakarta where melioidosis was very rarely reported, identification was a big challenge. *Burkholderia pseudomallei* was not identified in the early phase of the patient illness that might cause delay of appropriate infection management, which eventually might lead to patient death. Patient suffered from pneumonia, hypoalbuminemia, electrolyte imbalance, acute pancreatitis, post-operative *colo-cutaneus* fistula with secondary infection, acute kidney injury, melena, sepsis, and prolong hospitalization as the consequences of etiology misidentification. Prolong hospitalization, and immobilization also contributed to hospital-acquired pneumonia with bilateral lung effusion, which finally lead to acute respiratory distress syndrome (ARDS).

The isolated *B. pseudomallei* which susceptible to meropenem and amikacin only, and the presence of other pathogens (TABLE 3) altogether developed into infectious agents package which very difficult to combat. The physicians have struggled to stabilize patient condition, with the available resource. Unfortunately, antibiotic administration management was also challenging since empirical treatment in this case was prone to ineffective to diminish infection, then severe complications accompanied, that made infection even more difficult to be cured. It showed the dynamic of antibiotic treatment and the disease journey. It showed that patient had enduring prolong fever, infection that unresponsive to antibiotic treatment, and recurrent septic shocks before finally he passed away.

CONCLUSION

Lesson learned from this case is the crucial of diagnoses establishment and appropriate antibiotic treatment in melioidosis management. In addition, in the setting where resource are limited, a rapid diagnostic tool will help the clinicians to adjust the empirical treatment prior the culture result.

ACKNOWLEDGEMENTS

Authors are acknowledge all surgery and internal medicine residents from Dr. Sardjito General Hospital, Yogyakarta who help to treat the patient.

REFERENCES

1. Wiersinga WJ, Virk HS, Torres AG, Currie BJ, Peacoco SJ, Dance DAB, *et al.* Melioidosis. Nat Rev Dis Primers 2018; 4:17107.

https://doi.org/10.1038/nrdp.2017.107

- Cheng AC, Currie BJ. Melioidosis: epidemiology, pathophysiology, and management. Clin Microbiol Rev 2005; 18(2):383-416. h t t p s : // d o i . o r g / 1 0 . 1 1 2 8 / CMR.18.2.383-416.2005
- Crowe A, McMahon N, Currie BJ, Baird RW. Current antimicrobial susceptibility of first-episode melioidosis *Burkholderia pseudomallei* isolates from the Northern Territory, Australia. Int J Antimicrob Agents 2014; 44(2):160-2. h t t p s : // d o i . o r g / 1 0 . 1 0 1 6 / j . ijantimicag.2014.04.012
- 4. Lipsitz R, Garges S, Aurigemma R, Baccam P, Blaney DD, Cheng AC, *et al.* Workshop on treatment of and postexposure prophylaxis for *Burkholderia pseudomallei* and *B. mallei* infection, 2010. Emerg Infect Dis 2012; 18(12):e2. https://dx.doi.org/10.3201/

eid1812.120638

- 5. Currie BJ, Dance DA, Cheng AC. The global distribution of *Burkholderia pseudomallei* and melioidosis: an update. Trans R Soc Trop Med Hyg 2008; 102(Suppl 1):S1-4. https://dx.doi.org/10.1016/S0035-9203(08)70002-6
- Limmathurotsakul D, Golding N, Dance DA, Messina JP, Pigott DM, Moyes CL, et al. Predicted global distribution of Burkholderia pseudomallei and burden of melioidosis. Nat Microbiol 2016; 1(1):15008.

htpp://doi.org/10.1038/nmicrobiol.2015.8

 Hoffmaster AR, AuCoin D, Baccam P, Baggett HC, Baird R, Bhengsri S, *et al.* Melioidosis diagnostic workshop, 2013. Emerg Infect Dis 2015; 21(2):e141045

https://doi.org/10.3201/eid2102.141045

 Fisher DA, Harris PN. Melioidosis: refining management of a tropical time bomb. Lancet 2014; 383(9919): 762-4.

https://doi.org/10.1016/S0140-6736(13)62143-1

- 9. Anuntagool N, Naigowit P. Petkanchanapong V. Aramsri P. Panichakul T, Sirisinha S. Monoclonal antibody-based rapid identification of Burkholderia pseudomallei in blood culture fluid from patients with communityacquired septicaemia. Ι Med Microbiol 2000; 49(12):1075-8. https://doi.org/10.1099/0022-1317-49-12-1075
- 10. Chantratita N, Tandhavanant S, Wongsuvan G, Wuthiekanun V, Teerawattanasook N, Day NPJ,*et al.* Rapid detection of *Burkholderia pseudomallei* in blood cultures using a monoclonal antibody-based immunofluorescent assay. Am J Trop Med Hyg 2013; 89(5):971-2. https://doi.org/10.4269/ajtmh.13-0212
- Meumann 11. Chantratita N, E. Thanwisai A.Limmathurotsakul D, Wuthiekanun V, Wannapasni S, et al. Loop-mediated isothermal amplification method targeting the TTS1 gene cluster for detection of Burkholderia pseudomallei and diagnosis of melioidosis. J Clin Microbiol 2008; 46(2):568-73. https://doi.org/10.1128/JCM.01817-07
- 12. Jorakate P, Higdon M, Kaewpan A, Makprasert S, Yuenprakhon S, Tawisaid K, *et al.* Contribution of the BacT/Alert MB Mycobacterium bottle to bloodstream infection surveillance in Thailand: added yield for *Burkholderia pseudomallei*. J Clin Microbiol 2015; 53(3):910-4. https://doi.org/10.1128/JCM.02008-14
- 13. Dulsuk A, Paksanont S, Sangchankoom A, Ekchariyawat P, Phunpang R, Jutrakul Y, *et al.*

Validation of a monoclonal antibodybased immunofluorescent assay to detect *Burkholderia pseudomallei* in blood cultures. Trans R Soc Trop Med Hyg 2017; 110(11):670-2.

httpss://doi.org/10.1093/trstmh/trw079

- 14. Pongsunk S, Thirawattanasuk N, Piyasangthong N, Ekpo P. Rapid identification of *Burkholderia pseudomallei* in blood cultures by a monoclonal antibody assay. J Clin Microbiol 1999; 37(11):3662-7.
- 15. Woods KL, Boutthasavong L, NicFhogartaigh C, Lee SJ, Davong V, AuCoin DP, *et al.* Evaluation of a rapid diagnostic test for detection of *Burkholderia pseudomallei* in the Lao People's Democratic Republic. J Clin Microbiol 2018; 56(7):02002-17 https://doi.org/10.1128/JCM.02002-17
- 16. Karger A, Stock R, Ziller M, Elschner MC, Bettin B, Melzer F, et al. Rapid identification of Burkholderia mallei and Burkholderia pseudomallei by intact cell Matrix-assisted Laser Desorption/Ionisation Mass Spectrometric typing. BMC Microbiol 2012; 12:229

https://doi.org/10.1186/1471-2180-12-229

17. Peddayelachagiri BV, Paul S, Gogoi M, Sripathy MH, Batra HV. Evaluation of fimC and bdha based duplex PCR for specific identification and differentiation of *Burkholderia pseudomallei* from near-neighbor *Burkholderia* species. Int J Med Microbiol 2018; 308(2):271-8. https://doi.org/10.1016/j. ijmm.2017.11.007

- 18. Tellapragada C, Shaw T, D'Souza A, Eshwara VK, Mukhopadhyay C. Improved detection of *Burkholderia pseudomallei* from non-blood clinical specimens using enrichment culture and PCR: narrowing diagnostic gap in resource-constrained settings. Trop Med Int Health 2017; 22(7):866-70. https://doi.org/10.1111/tmi.12894
- 19. Wuthiekanun V. Dance D, Limmathurosakul D. Colony Burkholderia morphology of pseudomallei on different culture media. Bandung, Indonesia: Welcome trust MORU Tropical Health Network; 2017. p.1.
- 20. Kingsley PV, Arunkumar G, Tipre M, Leader M, Sathiakumar N. Pitfalls and optimal approaches to diagnose melioidosis. Asian Pac J Trop Med 2016; 9(6):515-24. https://doi.org/10.1016/j.
 - apjtm.2016.04.003