# Screening and Prediction of Potential Compounds from Virgin Olive Oil Acting on Proteins Associated with Cancer Disease

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### ABSTRACT

Virgin olive oil contains phenolic compounds that were potential for anti-inflammatory and cancer treatment. Computational biology is a beneficial method to understand how this compound can affect the biological process in humans. This research is conducted by the potential screening of VOO compounds, constructing the pharmacological network and enrichment, and docking simulation. The enrichment result showed that the EGFR, BRAF, MAPK1, CCND1, and MDM2 protein have multiple cancer contributions and related pathways. The docking simulation result showed that the interaction of EGFR-luteolin, BRAF-luteolin, MAPK1-luteolin, CCND1-apigenin, and MDM2-1-hydroxypinoresinol has the highest binding affinity. Further research with the in-vitro methods are required to check the reliable mechanisms of each compound to their protein target.

Keywords: Bioinformatics; Cancer; Molecular docking; Network pharmacology; Olive oils; Phenolics

### **INTRODUCTION**

The virgin olive oil (VOO) is the main product of olive trees (Olea europaea subs. Eureupaea). This tree is a symbolic species in the Mediterranean region. Genomic research with this tree is important for facilitating study in metabolisms, developmental and physiological process, which is the research could contribute to improving the economic values of this tree (Cruz et al., 2016). Extra VOO contains dominant phenolic compounds (Visioli and Bernardini, 2011), which those compounds have different classes, such as phenyl ethyl alcohols, steroids, phenolic acids, hydroxy-isochromans, lignans and, flavonoids. The phenolics (also known as polyphenols) from a plant have some benefits in the human body to prevent various diseases. They work to bind several proteins, which may lead to a specific disease, and they have also antioxidants activity against free-radical (Preedy and Watson, 2010). In studies, they can the in-vitro modulate intracellular signaling pathways; SO, V00 compounds, like hydroxytyrosol, tyrosol, and other minor compounds, have been the focus of research to see its effect in the biological process (Serreli and Deiana, 2018).

Recent studies in-vitro and in-vivo with VOO compounds have been done. The recent studies, hydroxytyrosol from VOO has been reported could induce cell cycle arrest and apoptosis on various

\*Corresponding author: Achmad Rodiansyah Email : ach.rodiansyah@gmail.com cells, such as colon cancer cells, bladder cancer and cholangiocarcinoma (Coccia *et al.*, 2016; Li *et al.*, 2014; López de las Hazas *et al.*, 2017). Pinoresinol may also probably have antitumor activity in breast cancer cells (López-Biedma *et al.*, 2016), and p-HPE-EDA can inhibit colon cancer cells with inhibition of AMP-activation protein kinase (AMPK) and cyclooxygenase-2 (COX-2) (Khanal *et al.*, 2011). In silico and in vitro study showed that oleuropein confirmed could inhibit mTOR, which is responsible for tumor cells' properties on breast cancer (Corominas-Faja *et al.*, 2018).

World health organization (WHO) mentioned that cancer which causes the death of about 9.6 million death in 2018, is a group of diseases with abnormal cell growth that can invade other organs. The mechanisms in this disease are very complicated. Researchers mention that this disease is mainly caused by a gene mutation affecting cell functions, carcinogenic chemical compounds, and an unhealthy lifestyle (Hassanpour and Dehghani, 2017). Genetic materials and proteins play an essential role in this group's diseases; with bioinformatics, it will more easily explore potential clinical applications and improve diagnosis, therapies, and cancer diseases prognosis (Wu et al., 2012). Computational network biology is a new research field that involves theory and applications to describe a molecule's interaction on living cells; this field also contributes to accelerating molecular biology, pharmacology, and genetics studies (Ni et al., 2018). This method is acceptable for integrating

No	PubChem ID	Compound Name
1	5280934	alpha-Linolenic acid
2	5280450	Linoleic acid
3	445639	Oleic acid
4	985	Palmitic acid
5	5281	Stearic acid
6	10393	Tyrosol (p-HPEA)
7	82755	Hydroxytyrosol (3,4-DHPEA)
8	131750845	1-(3-Methoxy-4-hydroxy)-phenyl-6,7-dihydroxy-isochroman
9	442831	1-Acetoxypinoresinol
10	13824420	1-Hydroxypinoresinol
11	131750844	1-Phenyl-6,7-dihydroxy-isochroman
12	124202093	3,4-DHPEA-EA
13	5280443	Apigenin
14	5280445	Luteolin
15	16681728	p-Hpea-eda
16	234817	Pinoresinol

Table I. Bioactive compounds in virgin olive oil

and analyzing big data; with this network model, the extensive information of the biological system could be more easily to be understanding (Ideker and Nussinov, 2017; Ma'ayan, 2011).

### **METHODOLOGY**

### Preparation of ligand and protein target

VOO bioactive compounds were collected from the PubChem database (https://pubchem. ncbi.nlm.nih.gov/) based on relevant literature (Alavi and Golmakani, 2017; Basiricò et al., 2019; Rupani, 2014; Waterman, 2007; Zalejska-Fiolka et al., 2015). Finally, 16 bioactive compounds used in this study were listed in Table I. The 3D structure and canonical SMILES of all compounds were downloaded from the PubChem database. Canonical SMILES compounds were used for identifying the potential target proteins on the Phrammapper web server (http://lilab-ecust.cn/ pharmmapper/) and Swiss Target Prediction web server (http://www. swisstargetprediction.ch/). Phrammapper web server is a platform for identifying the potential protein target from bioactive compounds with statistical method calculation include more than 7000 target pharmacophores (X. Liu et al., 2010; X. Wang et al., 2017). The specification for searching target proteins was set as Druggable Pharmacophore Models (v2017,16159) (Liang *et al.*, 2019). Like a Phrammapper webserver, SwissTargetPrediction is an accruable webserver to predict the target proteins of bioactive compounds (Gfeller et al., 2014); this web server has been updated for efficient prediction of protein targets (Daina et al., 2019). The result from Phammapper and SwissTargetPrediction webserver was saved as .csv format and used to construct network compound-protein interactions.

### Network construction and analysis

The network compound-protein interaction was constructed by Cytoscape software and its plugins (https://cytoscape.org/). Excel files from Pharmmapper and SwissTargetPrediction that contains compound-protein interaction data were imported to Cytoscape software with the menu "import network form file system". The compound table was selected as a "source node", and the protein target was selected as a "target node". The following analysis was used ClueGO Cytoscape's plugins to show the interpretation of list genes on the metabolical process (Bindea et al., 2009). The ClueGO plugin's setting performs as pathways with p-values  $\leq$  0.001, and the kappa score was set as 0.4. The Gene Ontology databases were used from the Kyoto Encyclopedia and Genomes of Genes (KEGG) and Wikipathways. The selected target proteins from the ClueGO enrichment were used for docking simulation for determining ligand-protein interaction.

# Protein Preparation and Molecular docking simulation

The ClueGO results relating to cancer diseases were selected as a target protein for molecular docking. The target protein used for molecular docking were retrieved from Protein Data Bank (https://www.rcsb.org/). The 3D structure of proteins used in this study was listed as follows: EGFR-PDB.id 6LUD (Kashima *et al.*, 2020), BRAF-PDB.id 6NSQ (Assadieskandar *et al.*,



Figure 1. The network interaction from Cytoscape of 16 compounds of VOO with their target proteins

2019), MDM2-PDB.id 3LNJ (M. Liu et al., 2010), MAPK/ERK-PDB.id 5NHH (Ward et al., 2017), and CCND1 6P6G (Guiley et al., 2019). Those target proteins were sterilized from waters and ligands using PyMOL software (available in https://pymol.org/2/). The ligands used for molecular docking simulation were converted to .pdbqt format using open babel on PyRx software (available in https://pyrx.sourceforge.io/), and the reverse docking simulation was performed using Vina Wizard in PyRx (Dallakyan and Olson, 2015; Trott and Olson, 2010). The docking results were visualized and evaluated using PyMOL software. The interaction between amino acid residues and ligands in 2D view were visualized using BIOVIA DiscoveryStudio 2019 software (available in https://www.3dsbiovia.com/).

# **RESULT AND DISCUSSION**

# Preparation of ligand and network data construction

Sixteen potential ligands from VOO were collected from PubChem, and the list of targets from Phrammapper protein and SwissTargetPrediction were downloaded. Each compound interacted with various proteins that may contribute to many functions on biological pathways to build the compoundproteins network. Network construction with Cytoscape software showed 1639 nodes and 4016 edges. A node represents the target proteins and VOO compounds, while an edge represents the interaction of compounds-target proteins on the biological pathways (Figure 1). The VOO compounds labeled with red nodes in Figure 1 interact with blue nodes labeled as target proteins. Luteolin, 1-Hydroxypinorisenol, apigenin, linoleic acid, pinoresinol, oleic acid, stearic acid, 1-Acetoxypinorisenol, palmitic acid, p-Hpea-eda, and alpha-linolenic acid are located on central of nodes target protein, and other compound's nodes are located on outside of central interaction. Table panel protein target interactions from the Cytoscape network were saved as an Excel file. All protein was filtering and grouping to determine their biological pathways using ClueGO Cytoscape's plugin.

# Network construction and analysis

ClueGO analysis with KEGG and gene ontologies database showed 26 nodes and 38 edges with three pathways (Figure 2a), and WikiPathways database 35 nodes and 59 edges with four pathways (Figure 2b). ClueGO's plugin pathways were listed in Table 2; this result showed that VOO compounds are associated with various pathways, especially in tumor and cancer. The VOO compounds interact with various proteins that work on several biological pathways and are interconnected with its pathway or other pathways. From the KEGG database and WikiPathways database enrichment, it has known that VOO compounds' target protein was probably related to cancer disease and tumor with a p-value  $\leq$  of 0.001. A significant test of the p-value is vital for decision-making. A very small p-value indicated that the hypothesis is probably correct (Panagiotakos, 2008). These enrichment pathways indicated that they have a high confidence value.

All pathways from ClueGO are related to each other indicated that the several nodes of the gene connect with all pathways in Figure 2 (marked with a red circle). Glioma is the one type of tumours disease that occurs in the Central



Figure 2. ClueGO grouping shows the biological pathway from the list of the genes from the network. (a) KEGG database; (b) Wikipathways database.

Table II. ClueGO enric	hment pathways with	KEGG pathways database	e and WikiPathways database

No	Source Database	Pathway	Colour	P-Value
1	KEGG ontologies	Prostate cancer	Avocado green	7.77 x 10-9
2	KEGG ontologies	Bladder cancer	Turquoise	4.13 x 10-6
3	KEGG ontologies	Glioma	Dull green	3.60 x 10-6
4	Wikipathways	Glioblastoma	Blue	7.66 x 10-8
5	Wikipathways	Bladder cancer	Turquoise	6.28 x 10-6
6	Wikipathways	Oncostatin M signalling pathway	Green	1.49 x 10-6
8	Wikipathways	Matrix metalloproteinases	Dull green	4.37 x 10-6

Nervous System (CNS) and spinal cord; this disease is a common type of primary malignant brain tumor derived from glial cells (Ernest and Sontheimer, 2009). Glioblastoma or glioblastoma multiforme (GBM) is a subclass of glioma disease (JOVČEVSKA *et al.*, 2013).

Oncostatin M (OSM) is an important pathway associated with the biological process and cellular responses. This pathway is vital for clinical and biomedical therapeutic on the human disease

(Dey *et al.*, 2013). OSM pathway has been reported associating with cancer cell's plasticity (Junk *et al.*, 2017); also contribute to breast tumor specifically mediated by OSMR $\beta$  (Underhill-Day and Heath, 2006) and prostate cancer (Godoy-Tundidor *et al.*, 2005). Prostate cancer and bladder cancer are double cancer primary cancer with high frequency reported; this data suggest that the patients diagnosed with bladder or prostate cancer should be followed second malignant urologic diagnosis

(Kinoshita *et al.*, 2004). Disturbing the OSM pathway may is potential for cancer cell treatment (Caffarel and Coleman, 2014; Stroeder *et al.*, 2018).

The other pathway, Matrix metalloproteinases (MMPs), is a group of enzymes responsible for the degradation of extracellular matrix protein during organogenesis and normal tissue replacement. This group of enzymes is also associated with oral cancer (Sorsa *et al.*, 2004). Based on ClueGO enrichment, it is shown that this pathway has a connection with bladder cancer and OSM. The interruption of this protein activity could lead to various diseases (Laronha and Caldeira, 2020).

From KEGG and Wikipathways database enrichment, five genes contribute to OCM, glioma, bladder, and prostate cancer pathways. Those genes are CCND1 encodes G1/S-specific cyclin-D1 protein, MAPK1 encodes dual specificity mitogenactivated protein kinase 1, MDM2 encodes E3 ubiquitin-protein ligase Mdm2, EGFR encodes epidermal growth factor receptor protein, BRAF encodes serine/threonine-protein kinase B-RAF. These proteins were chosen for molecular docking simulation docked with the compound from VOO as the ligand to know their interaction. They could be evaluated to be used as therapeutic compounds for treating cancer disease.

The CCND1 (Cyclin D1) gene has a function in the regulation of CDK kinase in the cell cycle; when this gene had the mutation and overexpressed, it can promote a various type of cancers disease in humans, such as breast cancer, endometrial cancer, colon cancer, and prostate cancer (Fu et al., 2004; Ikeda et al., 2013; Moreno-Bueno et al., 2003; Xu and Lin, 2018). This gene is also used as a biomarker in breast cancer (Lundberg et al., 2019). Cyclin D1 protein functions as cyclin-dependent-kinase (CDK) in a subunit of CDK4 or CDK6 to regulate the cell cycle from G1 to S phase transition. They act as apoptosis regulators interacting with tumor suppressor protein retinoblastoma (Rb) to interrupting the cell cycle. CCND1 also acts on chromatin recruitment, mitochondrial biogenesis, and DNA Damage Response (DDR) (Fu et al., 2004; Massagué, 2004; Pestell, 2013). When this gene act as an oncogene mutation. local caused bv point DNA rearrangements, or chromosomal translocation, then they could overexpress in a cell, and that cell will have rich of complex CDK-cyclin; so, it could stimulate the progression in cell cycles, stimulate tumorigenesis, and metastases even in the absence of growth factor (Fu et al., 2004; Hardin et al., 2012; Kim and Diehl, 2009).

The murine double minute 2 (Mdm2) gene encodes a protein acting as a negative regulator of the p53 tumor suppressor. These genes' overexpression occurs in many tumors (Iwakuma and Lozano, 2003; Senturk and Manfredi, 2012). Mdm2 most commonly associated with liposarcoma compared with breast cancer and bladder cancer in MD Anderson phase 1 clinic (Dembla et al., 2018), and Mdm2 amplification not in most tumor types, that is just in the small subset in types tumor (Kato et al., 2018). The Mdm2 interacts with p53 protein to make p53 inactive, and this mdm2-p53 interaction can cause apoptosis failure (Hardin et al., 2012). Activation of p53 protein is critical to protect the propagation cells from damaged DNA with oncogenic mutations and control the cell cycle (Moll and Petrenko, 2003). Blocking the Mdm2-p53 and Mdm-non p53 interaction is a promising cancer therapeutic strategy (Shangary and Wang, 2008; S. Wang et al., 2017).

BRAF encodes Raf protein and MAPK1/2 gene-encoded mitogen-activated protein kinase 1/2 or Extracellular Signal-Regulated Kinases (ERKs) protein which play on signal transduction in the MAPK signaling pathway. This protein is known as signal transduction which works on the Ras-Raf-MEK-ERK signaling for cell proliferation, cell differentiation, cell metastasis, cell survival, and apoptosis (Mansfield et al., 2018; Mebratu and Tesfaigzi, 2009; Thatcher, 2010). Activating this pathway starts with growth factor family protein like Epidermal Growth Factor (EGF) binds their receptor to phosphorylate Raf protein as downstream of Ras on the MAPK signaling pathway (Ursem et al., 2018). Altered of this signaling pathway or mutation in genes encoding the protein in this pathways have been reported correlated and detected in tumors and cancer disease, including in pituitary tumourigenesis, cervical cancer tissue, and significantly correlated on breast cancer with axillary lymph node metastasis (Suojun et al., 2012; Manousaridis et al., 2013 Jan 1; Li et al., 2015: 1; AACR Project GENIE 2017; Shao et al., 2018). The inhibitors with targetting on these kinases protein could treat malignant tumors; this inhibitor would be promising and challenging in future research (Liu et al., 2018; Suojun et al., 2012). National Comprehensive Cancer Network (NCCN) also recommending BRAF testing gene for diagnosis of colorectal cancer (CRC) (Ursem et al., 2018).

# **Molecular Docking Simulation**

Computational methods recently used in the biotechnology and pharmaceutical industries were



#### **Docking Simulation Score from VOO Compounds**



Figure 3. Docking result of the compounds with selected protein from KEGG and WikiPathways database

No	Ligand	PubChem Id	Protein	Score (kcal/mol)	Label
1	Osimertinib	71496458	EGFR	-8.3	Red
2	Vandetanib	3081361	EGFR	-8.1	Magenta
3	Encorafenib	50296675	BRAF	-8.7	Red
4	Vemurafenib	42611257	BRAF	-9.2	Magenta
5	AMG-232	58573469	MDM2	-6.2	Red
6	Idasanutlin	53358942	MDM2	-7.6	Magenta
7	Vemurafenib	42611257	MAPK	-8.6	Red
8	Encorafenib	50296675	MAPK	-7.4	Magenta
9	CDK4 inhibitor	5330797	CCND1	-8.6	Red

Table III. Docking score from control compounds

beneficial for drug discovery with high success and accuracy and reduced costs (Parenti and Rastelli, 2012; Suortti, 1997). Reverse docking is a promising drug prediction technique that acts on protein-related disease as inhibitors (Kharkar *et al.*, 2014); this method has excellent drug design and drug discovery success. The docking simulation result showed that VOO compounds have various docking scores to target proteins (Figure 3).

The bar chart in Figure 3 shows the docking simulation score from VOO compounds with their target proteins. The deep-blue chart is an EGFR protein; an orange chart is a BRAF protein; a yellow chart is a MAPK protein; a grey chart is an MDM2 protein, and a blue chart is a CCND1 protein. Luteolin from VOO has the highest docking score, -7.9 kcal/mol on EGFR protein and -9.5 kcal/mol on BRAF protein. 1-Hydroxypinoresinol has the highest docking score with -7.0 kcal/mol on MDM2 protein. Apigenin has the highest docking score with -6.7 kcal/mol on CCND1 protein; also, on MAPK protein, luteolin has the highest docking score of -9.0 kcal/mol, respectively (Figure 3). Compared to the control compound, only the docking score from Luteolin-BRAF and luteolin-MAPK were more stable. The docking score from control compounds was listed in Table III.



Figure 4. Molecular interaction VOO ligands with the target proteins. (a) EGFR; (b) BRAF; (c) MDM2; (d) MAPK; (e) CCND1.

Binding affinity is a critical aspect of drug design to produce potential ligands with high binding affinity to the target protein and low target binding affinity to non-targeted protein (Kairys *et al.*, 2019). The Gibbs energy of binding ( $\Delta$ Gb) or binding affinity is value to define a strong interaction between two molecules, so this is a crucial quantity for study molecule interaction (Vangone *et al.*, 2018). The type of bond on reverse docking is critical on binding affinity as a docking score, especially the hydrogen bond in ligand-protein interaction. The 2D visual interaction of the VOO ligands colored with green within their target protein can be seen in Figure 4, and the list of amino acid residues can be seen in Table IV.

The results from molecular docking showed that the most of the ligands occupied the vital region of the protein. This interaction could potentially disrupt the work of target protein, like preventing phosphorylation and inhibiting protein activation. A ligands' effectiveness to interfere with proteins' action can be predicted by the docking score from the inhibitor ligand to the protein. Many factors that contribute to binding affinity score as follow: the role of water, the existence of H-bonds, the different types of the bind of ligand-protein interaction include ionic interactions, Van der Waals and hydrogen bond interaction, hydrophobic interaction, Pi-alkyl bond, and Pi-sulfur bond; those factors must be considered to evaluate docking results (Arthur and Uzairu, 2019; Pantsar and Poso, 2018). However, not all parameters can be calculated on this docking simulation, so an in-vitro and in-vivo study must be carried to determine the reliable responses (Pintilie and Stefaniu, 2019).

The illustration of VOO compounds to inhibit the target protein can be seen in Figure 5. From this illustration, the VOO compounds probably can work on several fields on the cells, like on the extracellular-membrane layer, cytoplasm, and nucleus cells field. EGFR protein is located on membrane cell, MAPK1 and BRAF protein work on the cytoplasm, MDM2 and CCND1 protein work on the nucleus.

### CONCLUSION

The VOO compounds have multiple protein targets on various pathways, especially in cancers and tumors. Five proteins that act on multiple



Figure 5. The illustration of VOO compounds work on their target proteins in the cell (The illustration was created with BioRender. (Available at: https://biorender.com/)

No	Protein	VOO ligand	Amino acid residues	Domain UniProt
1	EGFR	luteolin	LEU718, GLY719, VAL726, ALA743,	Uniprot Id: P00533
			LYS 745 GLU762, LEU792, MET793,	Region: 712-972
			PR0794, GLY796, MET798, LEU844,	(Protein kinase
			THR854, ASP855	domain)
2	BRAF	luteolin	ILE463, VAL471, ALA481, VAL482,	Uniprot Id: P15056
			LYS483, GLU501, VAL504, LEU505,	Region: 457-717
			THR508, ILE513, LEU514, ILE527,	(Protein kinase
			THR529, GLN530, TRP531, CYS532,	domain)
			LEU567, PHE583, GLY593, ASP594,	
			PHE595	
3	MDM2	1-Hydroxypinoresinol	LEU54, LEU57, GLY58, ILE61, MET62,	Uniprot Id: Q00987
			TYR67, GLN72, HIS73, VAL93, VAL75,	Region: 1-110 (USP
			HIS96, ILE99, TYR100	interaction domain)
4	MAPK1	Luteolin	ILE31, GLY32, GLU33, GLY34, VAL39,	Uniprot Id: P28482
			ALA52, LYS54, ILE84, GLN105,	Region: 25-313
			ASP106, LEU107, MET108, GLU109,	(Protein kinase
			THR110, LYS114, LEU156, CYS166,	domain)
			ASP167	Region: 105-108
				(Inhibitor binding
				domain)
5	CCND1	Apigenin	ASN174, ILE178, ILE177, HIS181,	Uniprot Id: P24385
			VAL212, GLY214, LEU217, ARG218,	Region: 2-208
			PRO220, ASN222	(Interaction region)
				Region: 2-19 (Region
				for RPLP0 & TCF3)
				Region: 150-360
				(Region for TCF3)

Table IV. Amino acid residues from molecular docking simulation with the highest score and the information domain from UniProt

central pathways from KEGG and WikiPathways database were MAPK1, BRAF, EGFR, MDM2, and CCND1. The docking simulation showed that the luteolin compound was stable with EGFR protein and BRAF protein, with the docking score reaching about -7.9 kcal/mol and -9.5 kcal/mol, respectively; 1-hydroxypinoresinol was stable to interact with MDM2 protein, reaching a score of -7.0 kcal/mol; apigenin and stearic acid were stable to interact with CCND1 protein with docking score reaching -6.7 kcal/mol. Also, MAPK protein is stably interacting with luteolin, which has a docking score of -9.0 kcal/mol. Those compounds, especially luteolin, probably have the potential for therapeutic on various cancers and tumors. These docking simulation results also report that luteolin has a stronger binding affinity than the control compound for interfering with the BRAF and MAPK protein. The in-vitro and in-vivo study must be carried out to validate the specific response from ligands on proteins involved in cancer pathways.

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# REFERENCES

- AACR Project GENIE: Powering Precision Medicine through an International Consortium, 2017. Cancer Discov. 7, 818–831.
- Alavi, N., Golmakani, M.-T., 2017. Improving oxidative stability of virgin olive oil by addition of microalga Chlorella vulgaris biomass. J. Food Sci. Technol. 54, 2464– 2473.
- Arthur, D.E., Uzairu, A., 2019. Molecular docking studies on the interaction of NCI anticancer analogues with human Phosphatidylinositol 4,5-bisphosphate 3-kinase catalytic subunit. J. King Saud Univ. - Sci.
- Assadieskandar, A., Yu, C., Maisonneuve, P., Kurinov, I., Sicheri, F., Zhang, C., 2019. Rigidification Dramatically Improves Inhibitor Selectivity for RAF Kinases. ACS Med. Chem. Lett. 10, 1074–1080.
- Basiricò, L., Morera, P., Dipasquale, D., Bernini, R., Santi, L., Romani, A., Lacetera, N., Bernabucci, U., 2019. (-)-Epigallocatechin-3gallate and hydroxytyrosol improved antioxidative and anti-inflammatory responses in bovine mammary epithelial cells. animal 1–10.
- Bindea, G., Mlecnik, B., Hackl, H., Charoentong, P., Tosolini, M., Kirilovsky, A., Fridman, W.-H., Pagès, F., Trajanoski, Z., Galon, J., 2009.

ClueGO: a Cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks. Bioinformatics 25, 1091–1093.

- Caffarel, M.M., Coleman, N., 2014. Oncostatin M receptor is a novel therapeutic target in cervical squamous cell carcinoma: OSMR in cervical SCC. J. Pathol. 232, 386– 390.
- Coccia, A., Mosca, L., Puca, R., Mangino, G., Rossi, A., Lendaro, E., 2016. Extra-virgin olive oil phenols block cell cycle progression and modulate chemotherapeutic toxicity in bladder cancer cells. Oncol. Rep. 36, 3095– 3104.
- Corominas-Faja, B., Cuyàs, E., Lozano-Sánchez, J., Cufí, S., Verdura, S., Fernández-Arroyo, S., Borrás-Linares, I., Martin-Castillo, B., Martin, Á.G., Lupu, R., Nonell-Canals, A., Sanchez-Martinez, M., Micol, V., Joven, J., Segura-Carretero, A., Menendez, J.A., 2018. Extra-virgin olive oil contains a metaboloepigenetic inhibitor of cancer stem cells. Carcinogenesis 39, 601–613.
- Cruz, F., Julca, I., Gómez-Garrido, J., Loska, D., Marcet-Houben, M., Cano, E., Galán, B., Frias, L., Ribeca, P., Derdak, S., Gut, M., Sánchez-Fernández, M., García, J.L., Gut, I.G., Vargas, P., Alioto, T.S., Gabaldón, T., 2016. Genome sequence of the olive tree, Olea europaea. GigaScience 5, 29.
- Daina, A., Michielin, O., Zoete, V., 2019. SwissTargetPrediction: updated data and new features for efficient prediction of protein targets of small molecules. Nucleic Acids Res. 47, W357–W364.
- Dallakyan, S., Olson, A.J., 2015. Small-Molecule Library Screening by Docking with PyRx, in: Hempel, J.E., Williams, C.H., Hong, C.C. (Eds.), Chemical Biology. Springer New York, New York, NY, pp. 243–250.
- Dembla, V., Somaiah, N., Barata, P., Hess, K., Fu, S., Janku, F., Karp, D.D., Naing, A., Piha-Paul, S.A., Subbiah, V., Tsimberidou, A.M., Shaw, K., Meric-Bernstam, F., Hong, D.S., 2018.
  Prevalence of MDM2 amplification and coalterations in 523 advanced cancer patients in the MD Anderson phase 1 clinic. Oncotarget 9.
- Dey, G., Radhakrishnan, A., Syed, N., Thomas, J.K., Nadig, A., Srikumar, K., Mathur, P.P., Pandey, A., Lin, S.-K., Raju, R., Prasad, T.S.K., 2013. Signaling network of Oncostatin M pathway. J. Cell Commun. Signal. 7, 103–108.
- Ernest, N.J., Sontheimer, H., 2009. Glioma, in: Encyclopedia of Neuroscience. Elsevier, pp. 877–884.

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- Fu, M., Wang, C., Li, Z., Sakamaki, T., Pestell, R.G., 2004. Minireview: Cyclin D1: Normal and Abnormal Functions. Endocrinology 145, 5439–5447.
- Gfeller, D., Grosdidier, A., Wirth, M., Daina, A., Michielin, O., Zoete, V., 2014. SwissTargetPrediction: a web server for target prediction of bioactive small molecules. Nucleic Acids Res. 42, W32–W38.
- Godoy-Tundidor, S., Cavarretta, I.T.R., Fuchs, D., Fiechtl, M., Steiner, H., Friedbichler, K., Bartsch, G., Hobisch, A., Culig, Z., 2005. Interleukin-6 and oncostatin M stimulation of proliferation of prostate cancer 22Rv1 cells through the signaling pathways of p38 mitogen-activated protein kinase and phosphatidylinositol 3-kinase. The Prostate 64, 209–216.
- Guiley, K.Z., Stevenson, J.W., Lou, K., Barkovich, K.J., Kumarasamy, V., Wijeratne, T.U., Bunch, K.L., Tripathi, S., Knudsen, E.S., Witkiewicz, A.K., Shokat, K.M., Rubin, S.M., 2019. p27 Allosterically Activates Cyclin-Dependent Kinase 4 and Antagonizes Palbociclib Inhibition. Science 366.
- Hardin, J., Bertoni, G., Kleinsmith, L.J., Becker, W.M., 2012. Becker's world of the cell. Benjamin Cummings, Boston.
- Hassanpour, S.H., Dehghani, M., 2017. Review of cancer from perspective of molecular. J. Cancer Res. Pract. 4, 127–129.
- Ideker, T., Nussinov, R., 2017. Network approaches and applications in biology. PLOS Comput. Biol. 13, e1005771.
- Ikeda, Y., Oda, K., Hiraike-Wada, O., Koso, T., Miyasaka, A., Kashiyama, T., Tanikawa, M., Sone, K., Nagasaka, K., Maeda, D., Kawana, K., Nakagawa, S., Fukayama, M., Tetsu, O., Fujii, T., Yano, T., Kozuma, S., 2013. Cyclin D1 harboring the T286I mutation promotes oncogenic activation in endometrial cancer. Oncol. Rep. 30, 584–588.
- Iwakuma, T., Lozano, G., 2003. MDM2, An Introduction. Mol. Cancer Res. 1, 993–1000.
- JOVČEVSKA, I., KOČEVAR, N., KOMEL, R., 2013. Glioma and glioblastoma - how much do we (not) know? Mol. Clin. Oncol. 1, 935–941.
- Junk, D.J., Bryson, B.L., Smigiel, J.M., Parameswaran, N., Bartel, C.A., Jackson, M.W., 2017. Oncostatin M promotes cancer cell plasticity through cooperative STAT3-SMAD3 signaling. Oncogene 36, 4001–4013.
- Kairys, V., Baranauskiene, L., Kazlauskiene, M., Matulis, D., Kazlauskas, E., 2019. Binding affinity in drug design: experimental and computational techniques. Expert Opin. Drug Discov. 14, 755–768.

- Kashima, K., Kawauchi, H., Tanimura, H., Tachibana, Y., Chiba, T., Torizawa, T., Sakamoto, H., 2020. CH7233163 Overcomes Osimertinib -Resistant EGFR-Del19/T790M/C797S Mutation. Mol. Cancer Ther. 19, 2288–2297.
- Kato, S., Ross, J.S., Gay, L., Dayyani, F., Roszik, J., Subbiah, V., Kurzrock, R., 2018. Analysis of MDM2 Amplification: Next-Generation Sequencing of Patients With Diverse Malignancies. JCO Precis. Oncol. 1–14.
- Khanal, P., Oh, W.-K., Yun, H.J., Namgoong, G.M., Ahn, S.-G., Kwon, S.-M., Choi, H.-K., Choi, H.S., 2011. p-HPEA-EDA, a phenolic compound of virgin olive oil, activates AMP-activated protein kinase to inhibit carcinogenesis. Carcinogenesis 32, 545–553.
- Kharkar, P.S., Warrier, S., Gaud, R.S., 2014. Reverse docking: a powerful tool for drug repositioning and drug rescue. Future Med. Chem. 6, 333–342.
- Kim, J.K., Diehl, J.A., 2009. Nuclear cyclin D1: An oncogenic driver in human cancer. J. Cell. Physiol. 220, 292–296.
- Kinoshita, Y., Singh, A., Rovito, P.M., Wang, C.Y., Haas, G.P., 2004. Double Primary Cancers of the Prostate and Bladder: A Literature Review. Clin. Prostate Cancer 3, 83–86.
- Laronha, H., Caldeira, J., 2020. Structure and Function of Human Matrix Metalloproteinases. Cells 9.
- Li, S., Han, Z., Ma, Y., Song, R., Pei, T., Zheng, T., Wang, J., Xu, D., Fang, X., Jiang, H., Liu, L., 2014. Hydroxytyrosol inhibits cholangiocarcinoma tumor growth: An in vivo and in vitro study. Oncol. Rep. 31, 145– 152.
- Li, X.-W., Tuergan, M., Abulizi, G., 2015. Expression of MAPK1 in cervical cancer and effect of MAPK1 gene silencing on epithelialmesenchymal transition, invasion and metastasis. Asian Pac. J. Trop. Med. 8, 937– 943.
- Liang, J., Wang, M., Olounfeh, K.M., Zhao, N., Wang, S., Meng, F., 2019. Network pharmacologybased identifcation of potential targets of the flower of Trollius chinensis Bunge acting on anti-inflammatory effectss. Sci. Rep. 9.
- Liu, F., Yang, X., Geng, M., Huang, M., 2018. Targeting ERK, an Achilles' Heel of the MAPK pathway, in cancer therapy. Acta Pharm. Sin. B 8, 552–562.
- Liu, M., Pazgier, M., Li, Changqing, Yuan, W., Li, Chong, Lu, W., 2010. A left handed solution to peptide inhibition of the p53-MDM2 interaction. Angew. Chem. Int. Ed Engl. 49, 3649–3652.

- Liu, X., Ouyang, S., Yu, B., Liu, Y., Huang, K., Gong, J., Zheng, S., Li, Z., Li, H., Jiang, H., 2010. PharmMapper server: a web server for potential drug target identification using pharmacophore mapping approach. Nucleic Acids Res. 38, W609–W614.
- López de las Hazas, M.-C., Piñol, C., Macià, A., Motilva, M.-J., 2017. Hydroxytyrosol and the Colonic Metabolites Derived from Virgin Olive Oil Intake Induce Cell Cycle Arrest and Apoptosis in Colon Cancer Cells. J. Agric. Food Chem. 65, 6467–6476.
- López-Biedma, A., Sánchez-Quesada, C., Beltrán, G., Delgado-Rodríguez, M., Gaforio, J.J., 2016. Phytoestrogen (+)-pinoresinol exerts antitumor activity in breast cancer cells with different oestrogen receptor statuses. BMC Complement. Altern. Med. 16.
- Lundberg, A., Lindström, L.S., Li, J., Harrell, J.C., Darai-Ramqvist, E., Sifakis, E.G., Foukakis, T., Perou, C.M., Czene, K., Bergh, J., Tobin, N.P., 2019. The long-term prognostic and predictive capacity of cyclin D1 gene amplification in 2305 breast tumours. Breast Cancer Res. 21, 34.
- Ma'ayan, A., 2011. Introduction to Network Analysis in Systems Biology. Sci. Signal. 4, tr5-tr5.
- Manousaridis, I., Mavridou, S., Goerdt, S., Leverkus, M., Utikal, J., 2013. Cutaneous side effects of inhibitors of the RAS/RAF/MEK/ERK signalling pathway and their management [WWW Document]. J. Eur. Acad. Dermatol. Venereol.
- Mansfield, A.S., Dy, G.K., Ahn, M.-J., Adjei, A.A., 2018.
  48 New Targets for Therapy in Lung Cancer, in: Pass, H.I., Ball, D., Scagliotti, G.V. (Eds.), IASLC Thoracic Oncology (Second Edition). Content Repository Only!, Philadelphia, pp. 479-489.e6.
- Massagué, J., 2004. G1 cell-cycle control and cancer. Nature 432, 298–306.
- Mebratu, Y., Tesfaigzi, Y., 2009. How ERK1/2 Activation Controls Cell Proliferation and Cell Death Is Subcellular Localization the Answer? Cell Cycle Georget. Tex 8, 1168– 1175.
- Moll, U.M., Petrenko, O., 2003. The MDM2-p53 Interaction. Mol. Cancer Res. 1, 1001–1008.
- Moreno-Bueno, G., Rodríguez-Perales, S., Sánchez-Estévez, C., Hardisson, D., Sarrió, D., Prat, J., Cigudosa, J.C., Matias-Guiu, X., Palacios, J., 2003. Cyclin D1 gene (CCND1) mutations in endometrial cancer. Oncogene 22, 6115– 6118.
- Ni, Y., Müller, P., Wei, L., Ji, Y., 2018. Bayesian graphical models for computational

network biology. BMC Bioinformatics 19, 63.

- Panagiotakos, D.B., 2008. The Value of p-Value in Biomedical Research. Open Cardiovasc. Med. J. 2, 97–99.
- Pantsar, T., Poso, A., 2018. Binding Affinity via Docking: Fact and Fiction. Mol. J. Synth. Chem. Nat. Prod. Chem. 23.
- Parenti, M.D., Rastelli, G., 2012. Advances and applications of binding affinity prediction methods in drug discovery. Biotechnol. Adv. 30, 244–250.
- Pestell, R.G., 2013. New Roles of Cyclin D1. Am. J. Pathol. 183, 3–9.
- Pintilie, L., Stefaniu, A., 2019. *In Silico* Drug Design and Molecular Docking Studies of Some Quinolone Compound, in: Molecular Docking and Molecular Dynamics [Working Title]. IntechOpen.
- Preedy, V.R., Watson, R.R., 2010. Olives and olive oil in health and disease prevention. Elsevier, Amsterdam.
- Rupani, B., 2014. Enrichment of Olive Oil with Alpha Linolenic Acid Catalyzed by Lipase Mediated Trans-Esterification. Iran. J. Energy Environ. 5.
- Senturk, E., Manfredi, J.J., 2012. Mdm2 and Tumorigenesis. Genes Cancer 3, 192–198.
- Serreli, G., Deiana, M., 2018. Biological Relevance of Extra Virgin Olive Oil Polyphenols Metabolites. Antioxidants 7, 170.
- Shangary, S., Wang, S., 2008. Targeting the MDM2p53 Interaction for Cancer Therapy. Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res. 14, 5318–5324.
- Shao, G.-L., Wang, M.-C., Fan, X.-L., Zhong, L., Ji, S.-F., Sang, G., Wang, S., 2018. Correlation Between Raf/MEK/ERK Signaling Pathway and Clinicopathological Features and Prognosis for Patients With Breast Cancer Having Axillary Lymph Node Metastasis. Technol. Cancer Res. Treat. 17, 153303461775402.
- Sorsa, T., Tjäderhane, L., Salo, T., 2004. Matrix metalloproteinases (MMPs) in oral diseases. Oral Dis. 10, 311–318.
- Stroeder, R., Walch-Rückheim, B., Fischbach, J., Juhasz-Böss, I., Rübe, C., Solomayer, E.-F., Smola, S., 2018. Oncostatin M treatment increases the responsiveness toward cisplatin-based chemoradiotherapy in cervical cancer cells in a STAT3-dependent manner. Oncol. Lett. 16, 3351–3358.
- Suojun, Z., Feng, W., Dongsheng, G., Ting, L., 2012. Targeting Raf/MEK/ERK pathway in pituitary adenomas. Eur. J. Cancer 48, 389– 395.

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- Suortti, T., 1997. Coupled size-exclusion chromatography-anion-exchange chromatography in the analysis of poly- and oligosaccharides. J. Chromatogr. A 763, 331– 335.
- Thatcher, J.D., 2010. The Ras-MAPK Signal Transduction Pathway. Sci. Signal. 3, tr1– tr1.
- Trott, O., Olson, A.J., 2010. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading. J. Comput. Chem. 31, 455–461.
- Underhill-Day, N., Heath, J.K., 2006. Oncostatin M (OSM) Cytostasis of Breast Tumor Cells: Characterization of an OSM Receptor β-Specific Kernel. Cancer Res. 66, 10891-10901.
- Ursem, C., Atreya, C.E., Van Loon, K., 2018. Emerging treatment options for BRAFmutant colorectal cancer. Gastrointest. Cancer Targets Ther. 8, 13–23.
- Vangone, A., Schaarschmidt, J., Koukos, P., Geng, C., Citro, N., Trellet, M.E., Xue, L.C., Bonvin, A.M.J.J., 2018. Large-scale prediction of binding affinity in protein–small ligand complexes: the PRODIGY-LIG web server. Bioinformatics 35, 1585–1587.
- Visioli, F., Bernardini, E., 2011. Extra Virgin Olive Oil's Polyphenols: Biological Activities. Curr. Pharm. Des. 17, 786–804.
- Wang, S., Zhao, Y., Aguilar, A., Bernard, D., Yang, C.-Y., 2017. Targeting the MDM2–p53 Protein– Protein Interaction for New Cancer Therapy: Progress and Challenges. Cold Spring Harb. Perspect. Med. 7, a026245.
- Wang, X., Shen, Y., Wang, S., Li, S., Zhang, W., Liu, X., Lai, L., Pei, J., Li, H., 2017. PharmMapper

2017 update: a web server for potential drug target identification with a comprehensive target pharmacophore database. Nucleic Acids Res. 45, W356–W360.

- Ward, R.A., Bethel, P., Cook, C., Davies, E., Debreczeni, J.E., Fairley, G., Feron, L., Flemington, V., Graham, M.A., Greenwood, R., Griffin, N., Hanson, L., Hopcroft, P., Howard, T.D., Hudson, J., James, M., Jones, C.D., Jones, C.R., Lamont, S., Lewis, R., Lindsay, N., Roberts, K., Simpson, I., St-Gallay, S., Swallow, S., Tang, J., Tonge, M., Wang, Z., Zhai, B., 2017. Structure-Guided Discovery of Potent and Selective Inhibitors of ERK1/2 from a Modestly Active and Promiscuous Chemical Start Point. J. Med. Chem. 60, 3438–3450.
- Waterman, E., 2007. Active Components and Clinical Applications of Olive Oil. Olive Oil 12, 12.
- Wu, D., Rice, C.M., Wang, X., 2012. Cancer bioinformatics: A new approach to systems clinical medicine. BMC Bioinformatics 13, 71.
- Xu, J., Lin, D.I., 2018. Oncogenic c-terminal cyclin D1 (CCND1) mutations are enriched in endometrioid endometrial adenocarcinomas. PLoS ONE 13.
- Zalejska-Fiolka, J., Wielkoszyński, T., Rokicki, W., Dąbrowska, N., Strzelczyk, J.K., Kasperczyk, A., Owczarek, A., Błaszczyk, U., Kasperczyk, S., Stawiarska-Pięta, B., Birkner, E., Gamian, A., 2015. The Influence of  $\alpha$  -Lipoic Acid and Garlic Administration on Biomarkers of Oxidative Stress and Inflammation in Rabbits Exposed to Oxidized Nutrition Oils. BioMed Res. Int. 2015, 1–11.